

Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis¹⁻⁵

Michael F Holick

ABSTRACT

The purpose of this review is to put into perspective the many health benefits of vitamin D and the role of vitamin D deficiency in increasing the risk of many common and serious diseases, including some common cancers, type 1 diabetes, cardiovascular disease, and osteoporosis. Numerous epidemiologic studies suggest that exposure to sunlight, which enhances the production of vitamin D₃ in the skin, is important in preventing many chronic diseases. Because very few foods naturally contain vitamin D, sunlight supplies most of our vitamin D requirement. 25-Hydroxyvitamin D [25(OH)D] is the metabolite that should be measured in the blood to determine vitamin D status. Vitamin D deficiency is prevalent in infants who are solely breastfed and who do not receive vitamin D supplementation and in adults of all ages who have increased skin pigmentation or who always wear sun protection or limit their outdoor activities. Vitamin D deficiency is often misdiagnosed as fibromyalgia. A new dietary source of vitamin D is orange juice fortified with vitamin D. Studies in both human and animal models add strength to the hypothesis that the unrecognized epidemic of vitamin D deficiency worldwide is a contributing factor of many chronic debilitating diseases. Greater awareness of the insidious consequences of vitamin D deficiency is needed. Annual measurement of serum 25(OH)D is a reasonable approach to monitoring for vitamin D deficiency. The recommended adequate intakes for vitamin D are inadequate, and, in the absence of exposure to sunlight, a minimum of 1000 IU vitamin D/d is required to maintain a healthy concentration of 25(OH)D in the blood. *Am J Clin Nutr* 2004;79:362-71.

KEY WORDS Vitamin D, sunlight, 25-hydroxyvitamin D, cancer, bone health, diabetes

INTRODUCTION

Once our sun ignited, it began to emit enormous amounts of energy. This energy bombarded all of its satellite planets. The third planet from the Sun (ie, Earth) had a huge ocean and a small land mass. In the bubbling, organically rich tide pools, life began to evolve and became dependent on solar energy for its very existence. Early in evolution, organisms captured the sun's energy in the form of carbohydrates through the process of photosynthesis. As organisms evolved, they continued to make a wide variety of complex macromolecules, not only for the purpose of replication but also to sustain life's functions. The life forms took advantage of their ocean environment and became dependent on calcium for signal transduction and metabolic functions. In ad-

dition, calcium became an important component for organisms that developed exoskeletons. The use of calcium for structural scaffolding became critically important in the evolution of ocean-dwelling vertebrates. The plentiful calcium in the oceans provided the ideal element to incorporate into a collagen-based matrix that gave rise to the structurally rigid vertebrate skeleton. The development of the vertebrate endoskeleton not only provided an opportunity for organisms to grow in size but also gave organisms the opportunity to venture onto land. As vertebrate organisms left their ocean environment for a land-based existence, they needed to develop an efficient method of utilizing the calcium that was absorbed into plants from the calcium-rich soil environment. Remarkably, it was the sun's energy that was called on to promote the photosynthesis of vitamin D₃ in the skin of vertebrates that was responsible for enhancing the efficiency of intestinal calcium absorption (1).

Little is known about when vitamin D made its appearance on Earth and what its function was. However, it is known that some of the earliest phytoplankton and diatom life forms, including *Emiliania huxleyi*, which has existed in the oceans for > 750 million years and which has used calcium for its structural support (it is a coccolithophore), produced ergosterol (provitamin D₂). When exposed to simulated sunlight, the ergosterol in *E huxleyi* was converted to previtamin D₂ (which rapidly isomerized to vitamin D₂; 2). *Skeletonema menziesii*, a diatom that also contained ergosterol, converted it to previtamin D₂. Little is known about the biologic function of ergosterol, previtamin D₂, and vitamin D₂ in nonvertebrate species. It has been suggested that ergosterol and its photoproducts are an ideal sunscreens system because of their high absorption of ultraviolet radiation (1). Ergosterol, previtamin D₂, vitamin D₂, and their photoproducts efficiently absorb the ultraviolet radiation that is damaging to

¹ From the Vitamin D, Skin, and Bone Research Laboratory, Section of Endocrinology, Diabetes, and Nutrition, Department of Medicine, Boston University School of Medicine, Boston.

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⁵ Address reprint requests to MF Holick, Boston University School of Medicine, 715 Albany Street, M-1013, Boston, MA 02118-2394. E-mail: mfhlick@bu.edu.

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Because the production of previtamin D₃ in the skin is directly related to the number of UVB photons that are absorbed by 7-dehydrocholesterol, any process that either decreases the number of UVB photons entering the epidermis or decreases the

amount of 7-dehydrocholesterol in the skin will result in a significant reduction in or the complete elimination of vitamin D₃ production in the skin.

A heightened awareness of the role that excessive exposure to sunlight plays in increasing the risk of nonmelanoma skin cancer and wrinkles led to the widespread use of topical sunscreens. Sunscreens efficiently absorb UVB radiation and thus markedly diminish the total number of UVB photons that reach the 7-dehydrocholesterol in the skin's cells. When used properly (ie, 2 mg/cm² or 35 mL—ie, 1 oz—on the whole body one time), a sunscreen with a sun protection factor of 8 reduces cutaneous production of previtamin D₃ by > 95% (10, 11). The proper use of a sunscreen with a sun protection factor of 15 reduces the capacity > 99%. The facts that most sunscreen users apply as little as 18% and no more than 35–50% of the recommended amount of sunscreen, and they do tan indicate that they are making sufficient amounts of vitamin D₃ in their skin. The fact that they tan is a reflection of the fact that UVB penetrates the epidermis to stimulate the melanocytes and make vitamin D₃. Melanin is a natural sunscreen that evolved to protect humans from blistering solar radiation as they evolved in equatorial regions of the world. This skin pigment is an extremely effective sunscreen with absorption properties from the ultraviolet C (200–280 nm) into the visible range (> 700 nm), and it competes quite well with 7-dehydrocholesterol for UVB photons. Thus, people of color who have greater amounts of melanin in their epidermis than do whites are less efficient in producing vitamin D₃ than are whites (11, 12). A person with skin type 5/6 (dark skin, never develops a sunburn) requires 10–50 times the exposure to sunlight to produce the same amount of vitamin D₃ in their skin as does a white person with skin type 2 or 3 (12).

The stratospheric ozone layer is efficient in absorbing all solar radiation below 290 nm. However, the ozone layer also can absorb UVB radiation above 290 nm that is responsible for producing previtamin D₃ in the skin. The ultraviolet radiation that can be absorbed by 7-dehydrocholesterol has energies down to 315 nm. Thus, when the angle of the sunlight (zenith angle) reaching the Earth's surface is very oblique (ie, early morning, late afternoon, and winter), sunlight must pass through more ozone, which efficiently absorbs the previtamin D₃-producing UVB photons, and thus very few, if any, reach the earth's surface. Because the zenith angle is dependent on time of day, season of the year, and latitude, those factors have a dramatic effect on the cutaneous production of vitamin D₃ (13, 14). Below ≈35°, the zenith angle is more direct, and therefore previtamin D₃ synthesis can occur in the skin year-round. However, above 35° latitude, the angle of the sun is so oblique during the winter months that most, if not all, of the UVB photons below 315 nm are absorbed by the ozone layer, thereby either reducing or completely preventing the production of previtamin D₃ in the skin. For example, residents of Boston (42°N), Edmonton, Canada (52°N), and Bergen, Norway (61°N) cannot produce sufficient quantities of vitamin D₃ in their skin for 4, 5, and 6 mo, respectively. We have conducted studies around the globe that provide guidelines for when, where, and at what time of day vitamin D₃ can be produced in the skin (14; **Figure 2**).

SOURCES OF VITAMIN D

Very few foods naturally contain vitamin D. Cod liver oil and oily fish such as salmon, mackerel, and sardines are good sources. Eating oily fish at least 3–4 times/wk will help satisfy

the requirement for adequate intake. Some foods such as milk (100 IU/8 oz), orange juice (100 IU/8 oz), and some cereals and breads are fortified with vitamin D (11, 12, 15). The vitamin D content in milk is often less than the label proclaims it to be, and thus the contribution of vitamin D from the diet is highly variable. To satisfy the body's requirement for vitamin D, most humans obtain it from casual exposure to sunlight. During the spring, summer, and fall, enough vitamin D₃ is produced in the skin to be stored in the body fat, and it can be mobilized during winter months when little, if any, vitamin D₃ is produced in the skin.

The skin has a large capacity to produce vitamin D₃. Blood concentrations of vitamin D₃ were compared in healthy young and middle-aged adults who were exposed to simulated sunlight that was equivalent to being on a sunny beach and obtaining enough sun to cause a slight pinkness to the skin (1 minimal erythema dose) and who took an oral dose of vitamin D₂. The exposure was equivalent to an oral dose of ≈20 000 IU vitamin D₂ (9; **Figure 3**). Although aging decreases the amount of 7-dehydrocholesterol produced in the skin by as much as 75% by the age of 70 y (16, 17), the skin has such a large capacity to make vitamin D₃ that even elderly exposed to sunlight can achieve increased blood concentrations of vitamin D₃ and 25(OH)D (17–19).

CAUSES AND CONSEQUENCES OF VITAMIN D DEFICIENCY

It is estimated that 10 million households in the United States have a reptile as a pet. In their natural environment, reptiles are often exposed to sunlight. They are vertebrates, and, like humans, they require a source of calcium and vitamin D. Iguanas are at particular risk of severe vitamin D deficiency because they are herbivores that, as pets, often are fed a steady diet of lettuce and because they are housed in glass enclosures with a light source that is devoid of UVB transmission. Lettuce contains very little calcium and no vitamin D, and thus iguanas and other vertebrates who do not receive an adequate amount of calcium and vitamin D develop the metabolic bone diseases osteoporosis and osteomalacia that result in fractures and ultimately death. Most reptile owners are aware of the need not only to provide their precious pets with a commercial source of calcium supplementation, but also to provide them with a light that emits UVB radiation similar to sunlight so that the animals can produce vitamin D₃ in their skin.

Humans are no different. They need an adequate source of calcium and vitamin D. Without vitamin D, the small intestine absorbs no more than 10–15% of dietary calcium. In a person with vitamin D sufficiency, the small intestine absorbs, on average, 30% of dietary calcium; during growth, lactation, and pregnancy, the efficiency increases to 80%. Vitamin D deficiency during bone development and growth causes the bone-deforming disease rickets. In adults bone growth stops and bone remodeling continues. Vitamin D deficiency in adults causes secondary hyperparathyroidism that can precipitate and exacerbate osteoporosis (2, 9, 11). The secondary hyperparathyroidism associated with vitamin D deficiency often maintains the serum calcium concentration within the normal range, but it causes a loss of phosphorus in the urine. This loss results in inadequate serum calcium × phosphorus to promote mineralization of the osteoid in the bone, which in turn results in osteomalacia, ie, nonmineralization of the collagen matrix. Because the nonmineralized matrix cannot provide structural support, the risk of fracture is greater.

How common is vitamin D deficiency? Surprisingly, it has made a resurgence in neonates and young children, in part be-

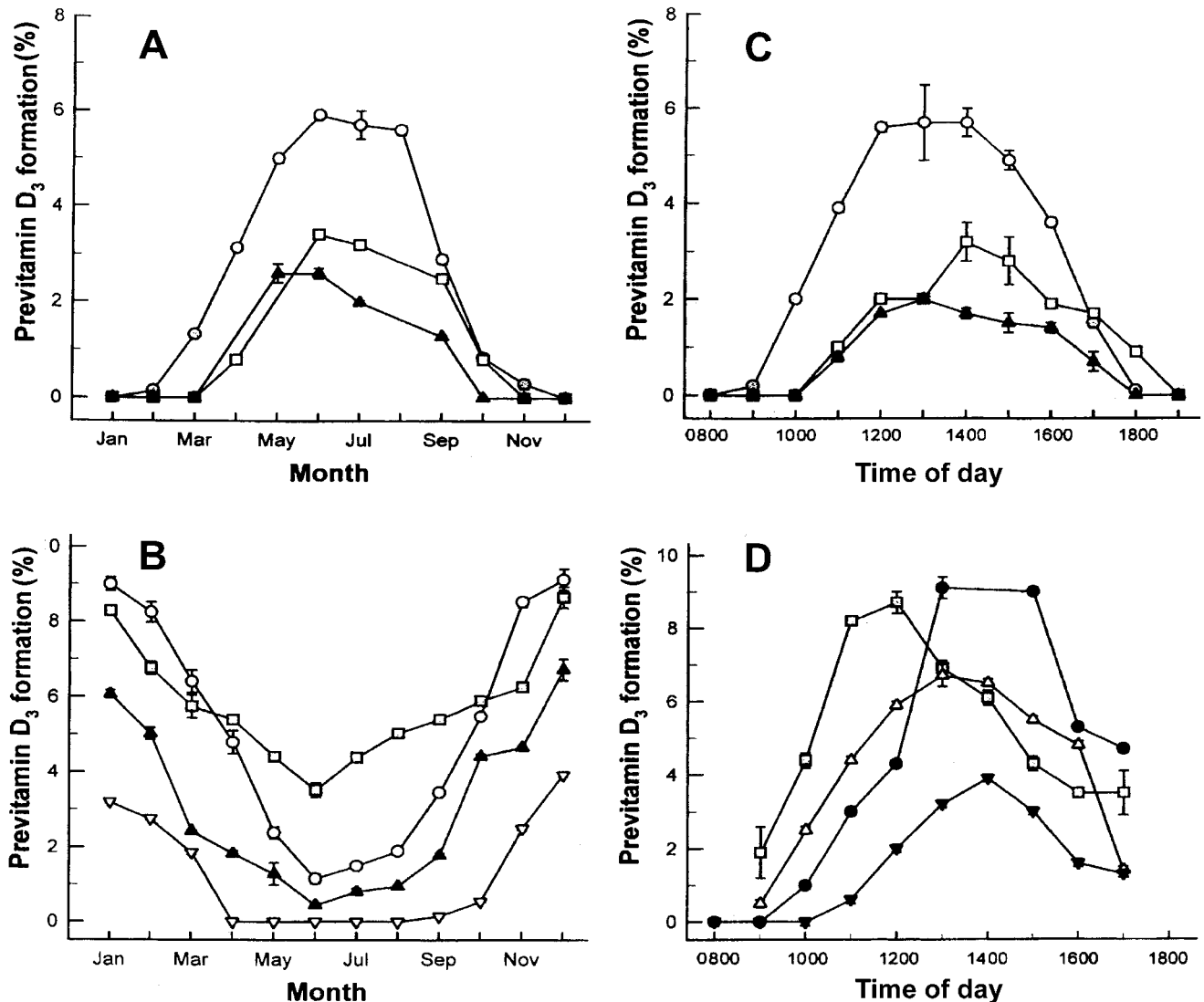


FIGURE 2. Influence of season, time of day, and latitude on the synthesis of previtamin D₃ in the northern (A and C: Boston, O; Edmonton, □; Bergen, ▲) and southern (B: Buenos Aires, O; Johannesburg, □; Cape Town, ▲; Ushuala, ▽; D: Buenos Aires, ●; Johannesburg, □; Cape Town, △; Ushuala, ▼) hemispheres. The hour indicated in C and D is the end of the 1-h exposure time in July and January, respectively. Adapted with permission (14).

cause of the campaign to encourage all women to provide all of their infants' nutrition through breastfeeding. Because there is very little, if any, vitamin D in human milk, infants, especially infants of women of color, are at high risk of developing vitamin D deficiency and rickets if they are not given a vitamin D supplement (20, 21).

The elderly are at risk for vitamin D deficiency because of poor dietary vitamin D intake and decreased exposure to sunlight. We observed that 30%, 42%, and 84% of free-living white, Hispanic, and black elderly were vitamin D deficient [25(OH)D < 50 nmol/L] at the end of August in Boston (9). It has always been assumed that young and middle-aged adults are not at risk of vitamin D deficiency because of their outdoor activities and dietary intake. However, it was recently recognized that 42% of African American women aged 15–49 y throughout the United States were vitamin D deficient [25(OH)D < 40 nmol/L] at the end of the winter (22). Hard-working young and middle-aged adults who very seldom spend any time outdoors or always wear sun protection outdoors are also at high risk

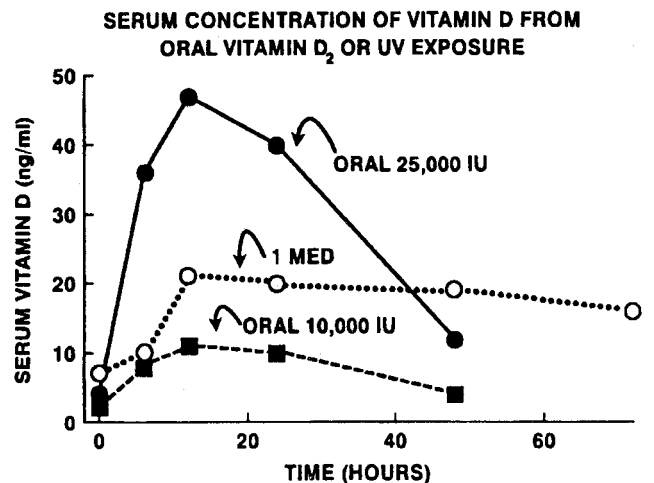


FIGURE 3. Serum vitamin D concentrations after a whole-body exposure to 1 minimal erythral dose (MED) of simulated sunlight in a tanning bed and after a single oral dose of either 10 000 or 25 000 IU vitamin D₂. UV, ultraviolet. Reproduced with permission (9).

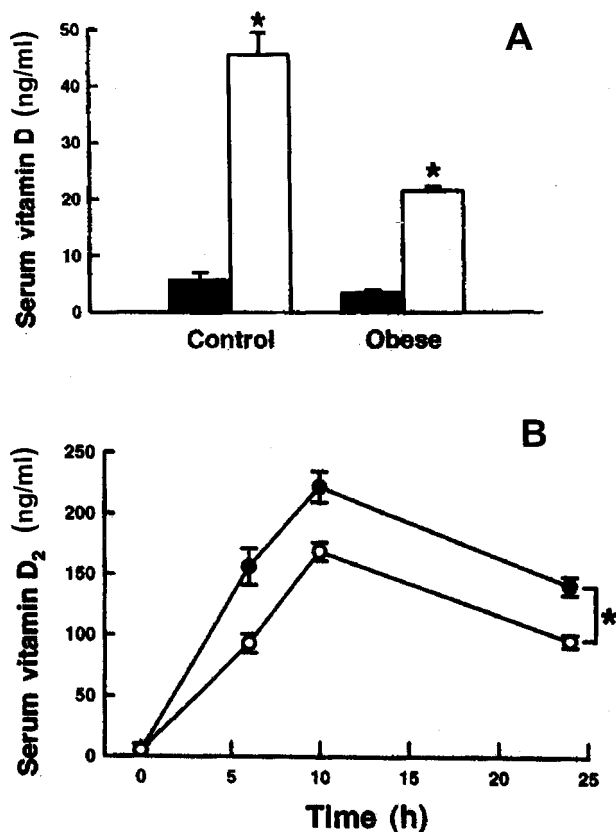


FIGURE 4. A: Mean (\pm SEM) serum vitamin D₃ concentrations before (■) and 24 h after (□) whole-body irradiation (27 mJ/cm²) with ultraviolet B radiation. The response of the obese subjects was attenuated when compared with that of the control group. There was a significant time-by-group interaction, $P = 0.003$. *Significantly different from preradiation values ($P < 0.05$). B: Mean (\pm SEM) serum vitamin D₂ concentrations in the control (●) and obese (○) groups before and 25 h after oral intake of vitamin D₂ (50 000 IU, or 1.25 mg). *Significant time and group effects by ANOVA ($P < 0.05$) but no significant time-by-group interaction. The difference in peak concentrations between obese and nonobese control subjects was not significant. Reproduced with permission (25).

of vitamin D deficiency. We observed that 32% of healthy adults 18–29 y of age were vitamin D deficient [$25(\text{OH})\text{D} < 50 \text{ nmol/L}$] at the end of the winter in Boston (23).

Obesity is often associated with vitamin D deficiency (24). It is now recognized that, whether vitamin D is ingested in the diet or obtained from exposure to sunlight, it is efficiently deposited in the large body fat stores and is not bioavailable (25; **Figure 4**). This is probably the reason that obese persons are chronically vitamin D deficient.

Vitamin D deficiency often goes undiagnosed or, worse, is misdiagnosed (9, 26–29). There are 3 reasons for this. First, it is believed that either exposure to sunlight or dietary intake of vitamin D is adequate, and, therefore, that Americans and Europeans are not at risk of vitamin D deficiency. Second, physicians who perform routine blood work-ups often obtain a blood calcium value. If they find it to be normal, they assume that the patient is vitamin D sufficient, which is not correct. Third, many physicians erroneously order an analysis for the active form of vitamin D, 1,25-dihydroxyvitamin D [$1,25(\text{OH})_2\text{D}$], to determine the vitamin D status of a patient. Unfortunately, $1,25(\text{OH})_2\text{D}$ not only is not a measure of vitamin D status, but its

measurement also can mislead the physician into thinking that the patient is vitamin D sufficient. The reason for this is that, as a person becomes vitamin D-deficient, there is an increase in the concentration of parathyroid hormone (PTH), which increases the renal production of $1,25(\text{OH})_2\text{D}$, the circulating concentrations of which often become normal or even elevated (9).

The vitamin D metabolite that should be measured to determine vitamin D status is $25(\text{OH})\text{D}$, which is the major circulating form of vitamin D, circulating at 1000 times the concentration of $1,25(\text{OH})_2\text{D}$ and having a half-life of ≈ 2 wk (2, 9, 11). As a person becomes vitamin D-deficient, there is a decrease in the efficiency of intestinal calcium absorption. The ionized calcium concentrations begin to drop; this decrease is immediately recognized by the calcium sensor in the parathyroid glands, which increases the production of PTH (30). PTH compensates for the decrease in intestinal calcium absorption by increasing the mobilization of calcium stores from the skeleton and by increasing tubular reabsorption of calcium in the kidney (31, 32).

NONSKELETAL CONSEQUENCES OF VITAMIN D DEFICIENCY

It has long been recognized that people who live at higher latitudes face an increased risk of many chronic diseases, including common cancers (33–39), multiple sclerosis (39, 40), and hypertension (41). As early as 1941, Apperly (37) observed that people living at higher latitudes, eg, Massachusetts and New Hampshire, had a higher risk of dying of the most common cancers than did people living in the South, eg, Georgia and South Carolina. In 1979, Rostand (41) reported that people living at higher latitudes in both the United States and Europe were at higher risk of hypertension. In the late 1980s and early 1990s, several investigators reported increased risks of dying of colon, prostate, and breast cancer in people living at higher latitudes in both the United States and Europe (33–35). Grant (42) reported that $\geq 25\%$ of the deaths due to breast cancer in women in Europe could be attributed to the women's lack of UVB from exposure to sunlight. Both men and women are at higher risk of dying of cancer if they have minimum exposure to sunlight (38; **Figure 5A and B**). In a retrospective study, Ahonen et al (44) reported that men on average begin to develop prostate cancer by the age of 52 y, whereas men exposed to more sunlight throughout their lives did not begin developing prostate cancer until 3–5 y later.

VITAMIN D METABOLISM AND NONCALCEMIC FUNCTIONS

It has been known for > 30 y that vitamin D₃ made in the skin or coming from the diet requires 2 obligate hydroxylations, first in the liver and then in the kidney, to create the active form of vitamin D, $1,25(\text{OH})_2\text{D}$ (**Figure 1**). $1,25(\text{OH})_2\text{D}$ interacts with its nuclear receptor in the intestine, bone, and kidney to regulate calcium and bone metabolism (9, 31, 45).

Most tissues and cells in the body, including heart, stomach, pancreas, brain, skin, gonads, and activated T and B lymphocytes, have nuclear receptors for $1,25(\text{OH})_2\text{D}$, called vitamin D receptors (46–48). Thus, it is not at all surprising that $1,25(\text{OH})_2\text{D}$ has a multitude of biologic effects that are noncalceimic in nature (9, 31, 45).

One of the most intriguing important and unappreciated biologic functions of $1,25(\text{OH})_2\text{D}$ is its ability to down-regulate hyperproliferative cell growth (9, 31, 49). Normal and cancer cells that have

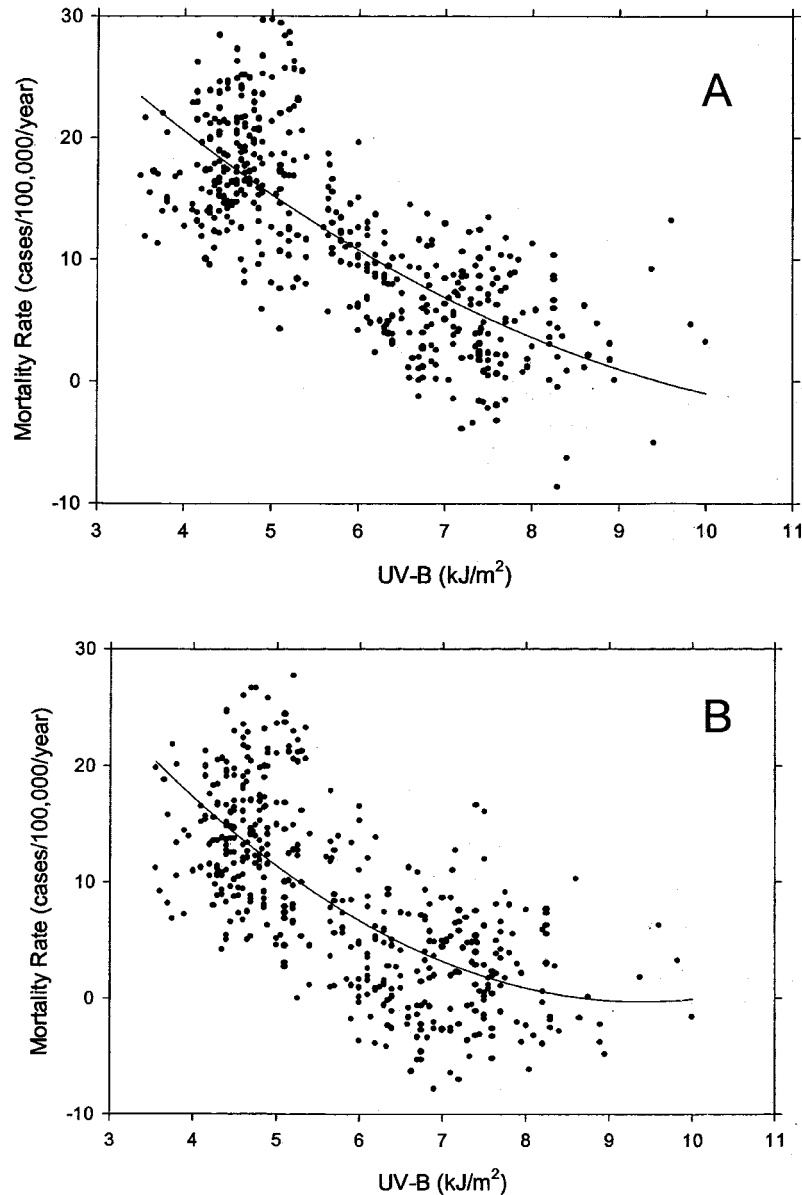


FIGURE 5. A: Premature mortality due to cancer in white females, as determined on the basis of the July 1992 DNA-weighted ultraviolet B (UV-B) radiation by use of a total ozone mapping spectrometer. B: Premature mortality due to cancer in white males in the United States from 1970 through 1994, as determined on the basis of the July 1992 DNA-weighted UV-B radiation. Reproduced by permission of Wiley-Liss, Inc, a subsidiary of John Wiley & Sons, Inc (43). Copyright (2002) American Cancer Society.

a vitamin D receptor often respond to $1,25(\text{OH})_2\text{D}$ by decreasing their proliferation and enhancing their maturation. This was the rationale for using $1,25(\text{OH})_2\text{D}_3$ and its analogs to treat the common hyperproliferative skin disorder psoriasis (50, 51).

Vitamin D receptors are present in activated T and B lymphocytes and in activated macrophages. The most common autoimmune diseases, including type 1 diabetes, rheumatoid arthritis, and multiple sclerosis, have all been successfully prevented in models using mice that were prone to these diseases if they received $1,25(\text{OH})_2\text{D}_3$ early in life (45, 52–55).

When nonobese diabetic mice, who typically develop type 1 diabetes, received $1,25(\text{OH})_2\text{D}_3$ throughout their life, their risk of developing type 1 diabetes was reduced by 80% (52, 55). This

is in good agreement with the recent observation by Hypponen et al (56) that children receiving 2000 IU vitamin D from age 1 y on decreased their risk of getting type 1 diabetes by 80%.

Krause et al (57) reported that hypertensive patients exposed to UVB radiation for 3 mo had a $> 180\%$ increase in circulating concentrations of $25(\text{OH})\text{D}$ and a 6 mm Hg decrease in their diastolic and systolic blood pressures, results similar to those expected if the patients had received a blood pressure medication (Figure 6). A similar group of patients who were exposed to ultraviolet A radiation and whose circulating concentrations of $25(\text{OH})\text{D}$ did not increase continued to be hypertensive throughout the 3-mo study. The exact mechanism by which UVB radiation returned the blood pressure to normal [presumably due to

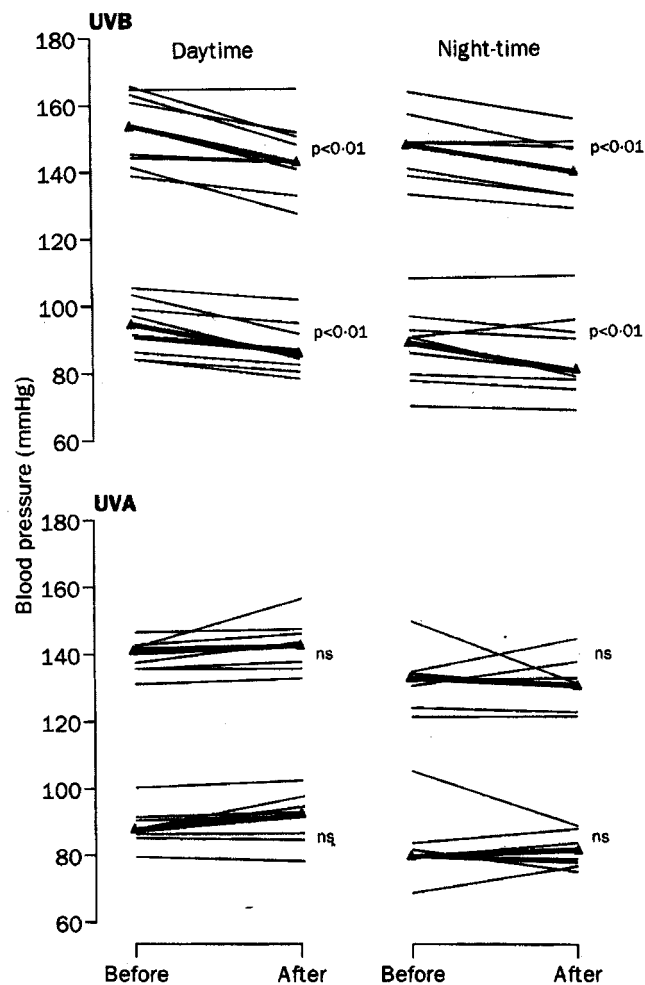


FIGURE 6. Effect of ultraviolet B (UVB) and ultraviolet A (UVA) radiation from a tanning bed on ambulatory daytime and nighttime blood pressure in hypertensive adults before and after exposure to tanning bed radiation 3 times/wk for 3 mo. The daytime and nighttime blood pressures after UVB tanning bed radiation were significantly ($P < 0.01$) different from those before irradiation. The mean is indicated by the thick line. Reproduced with permission from Elsevier (57).

increased blood concentrations of $25(\text{OH})\text{D}$ in these hypertensive adults is not well understood, but the observation by Li et al (58) sheds some light on the question. They observed in a mouse model that $1,25(\text{OH})_2\text{D}$ is effective in down-regulating renin and angiotensin and thereby decreasing blood pressure.

THE CANCER-VITAMIN D CONNECTION

Because an increased risk of vitamin D deficiency is one of the well-documented effects of living at higher latitudes on human health, it was reasonable to suggest that both living at higher latitudes and an increased risk of common diseases were associated with a decrease in the synthesis of vitamin D_3 in the skin. It was intuitively obvious to many, on the basis of new information about vitamin D metabolism and action, that increased exposure to sunlight at lower latitudes would increase blood concentrations of $25(\text{OH})\text{D}_3$, which could be activated in the kidney to $1,25(\text{OH})_2\text{D}_3$. Because $1,25(\text{OH})_2\text{D}_3$ is extremely potent in inhibiting cancer cell growth, this all seemed to make sense. Unfortunately, it was also well known that the renal production

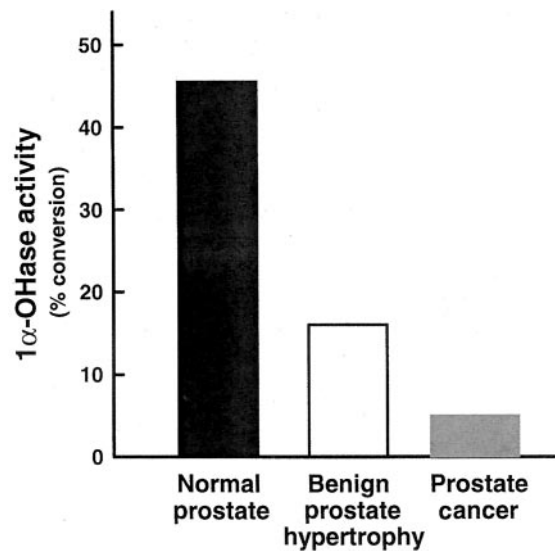


FIGURE 7. Measurement of 25-hydroxyvitamin D-1 α -hydroxylase (1α -OHase) activity in prostate cells that were either normal (■), benign but hypertrophied (□), or cancerous (▒). The activity was measured by incubating the cells with [^3H]25-hydroxyvitamin D_3 and measuring the percentage of conversion to [^3H]1,25-dihydroxyvitamin D_3 after 24 h.

of $1,25(\text{OH})_2\text{D}$ was tightly regulated by PTH, calcium, and phosphorus (31). Indeed, neither increased exposure to sunlight nor increased oral intake of vitamin D raised blood concentrations of $1,25(\text{OH})_2\text{D}$ (59–61). Thus, the question remained: how was it that increased exposure to sunlight was related, presumably by increasing the production of vitamin D_3 in the skin, to a decreased risk of many common cancers and other chronic diseases?

It had always been assumed that the kidney was the sole source for the body's production of $1,25(\text{OH})_2\text{D}$. This was based on many observations in animals and in humans whereby, in the absence of any renal function, there were little if any circulating concentrations of $1,25(\text{OH})_2\text{D}$. It had been reported that the placenta, epidermal cells, and bone cells could produce $1,25(\text{OH})_2\text{D}$, but the physiologic relevance of these observations was not well understood (62, 63). In 1985, Schwartz et al (64) reported that cultured prostate cancer cells expressed the enzymatic machinery to convert $25(\text{OH})\text{D}$ to $1,25(\text{OH})_2\text{D}$ (Figure 7). Since that observation, it has been shown that a wide variety of normal tissues as well as various cancer cells, including colon cancer, breast cancer, and lung cancer, all have the ability to make $1,25(\text{OH})_2\text{D}$ (65–67).

Thus, it is reasonable to conclude that increased exposure to sunlight or increased intake of vitamin D leads to higher circulating concentrations ($> 80 \text{ nmol/L}$) of $25(\text{OH})\text{D}$. $25(\text{OH})\text{D}$ acts as a substrate for the 25-hydroxyvitamin D-1-hydroxylase in various tissues, including colon, breast, and lung. These tissues can produce $1,25(\text{OH})_2\text{D}$, which acts in an autocrine fashion to regulate cell growth and decrease proliferative activity (Figure 8). It can also induce apoptosis when called on. Thus, $1,25(\text{OH})_2\text{D}_3$ can effectively manipulate cell growth and maintain it in a normal proliferative state under most circumstances. Once it accomplishes this, it induces the 25-hydroxyvitamin D-24-hydroxylase, which hydroxylates the $1,25(\text{OH})_2\text{D}$ in the side chain on carbons 23 and 24, and this results in cleavage between carbons 23 and 24 that forms the water-soluble, biologically inert calcitroic acid (31, 68; Figure 1). This is the likely explanation for why patients with renal failure develop a

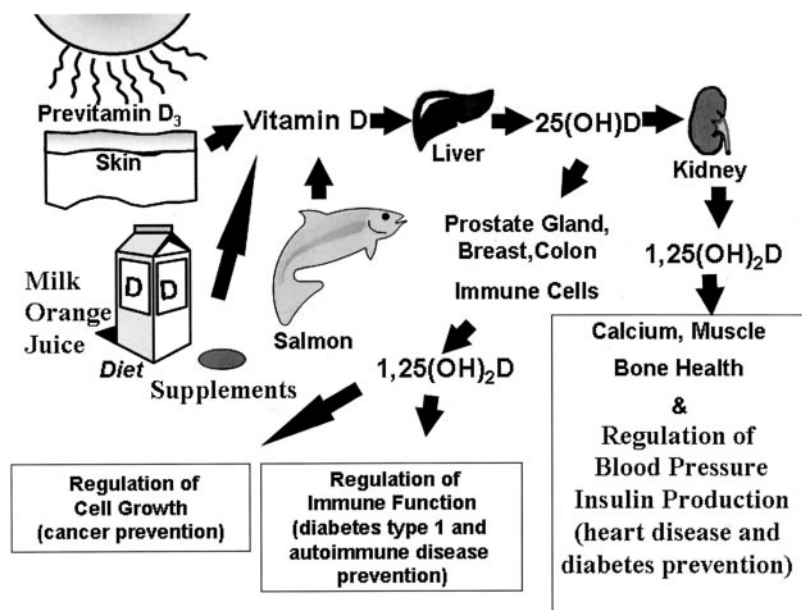


FIGURE 8. Schematic representation of the multitude of other potential physiologic actions of vitamin D with respect to cardiovascular health, cancer prevention, regulation of immune function, and decreased risk of autoimmune diseases. 25(OH)D, 25-hydroxyvitamin D; 1,25(OH)₂D, 1,25-dihydroxyvitamin D.

1,25(OH)₂D deficiency that results in secondary hyperparathyroidism and renal osteodystrophy.

CONCLUSIONS

The Institute of Medicine reported in 1997 that the recommended vitamin D intake was inadequate for adults over the age of 50 y (69). They recommended that the adequate intake for children and adults up to the age of 50 be 200 IU vitamin D/d. However, adults aged 50–70 y and ≥ 70 y required 400 and 600 IU vitamin D/d, respectively. As noted in Heaney's McCollum Award presentation (70) and as indicated in a considerable number of published reports, including that of Heaney et al (71), the new recommendations are totally inadequate, especially if a person has no exposure to sunlight. Without exposure to sunlight, a minimum of 1000 IU vitamin D/d is required. We gave healthy young and middle-aged adults 1000 IU vitamin D/d in orange juice from March through May. Their 25(OH)D concentrations increased by 150%, and what is considered to be a healthy 25(OH)D concentration, ie, 78–100 nmol/L (30–40 ng/mL), was maintained. Those adults receiving orange juice not fortified with vitamin D increased their blood concentration of 25(OH)D by 45%. This was due to their casual exposure to sunlight in the spring (15; **Figure 9**).

The easiest method of correcting vitamin D deficiency is to give the patient one pill that contains 50 000 IU vitamin D once a week for 8 wk (71). This will usually increase the 25(OH)D concentration to > 50 nmol/L (20 ng/mL; **Figure 10**). If not, the vitamin D “tank” may still not be full, and another 8-wk course of therapy usually corrects the vitamin D deficiency (67; **Figure 10**). One should suspect a fat-malabsorption problem or poor compliance if the 25(OH)D concentration does not increase by > 25% after these treatments. Exposure to sunlight or a tanning bed will correct vitamin D deficiency in patients with severe intestinal fat-malabsorption syndrome (72).

Why should we care about vitamin D deficiency? It is insidious and has both short- and long-term consequences. Infants and young children who are vitamin D deficient may be imprinted for the rest of their lives with increased risks of type 1 diabetes, multiple sclerosis, rheumatoid arthritis, and many common cancers (**Figure 8**). Adults are at increased risk of common cancers and cardiovascular disease. Recently, it has been reported that

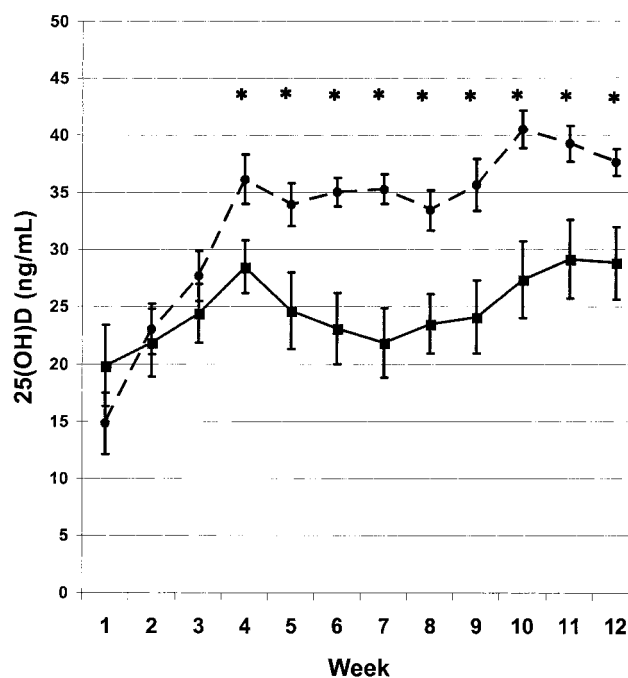


FIGURE 9. Mean (± SEM) weekly 25-hydroxyvitamin D [25(OH)D] concentrations in healthy adults ingesting orange juice fortified with vitamin D (1000 IU · 8 oz⁻¹ · d⁻¹, ●) or unfortified orange juice (■). **P* ≤ 0.01. Reproduced with permission (15).

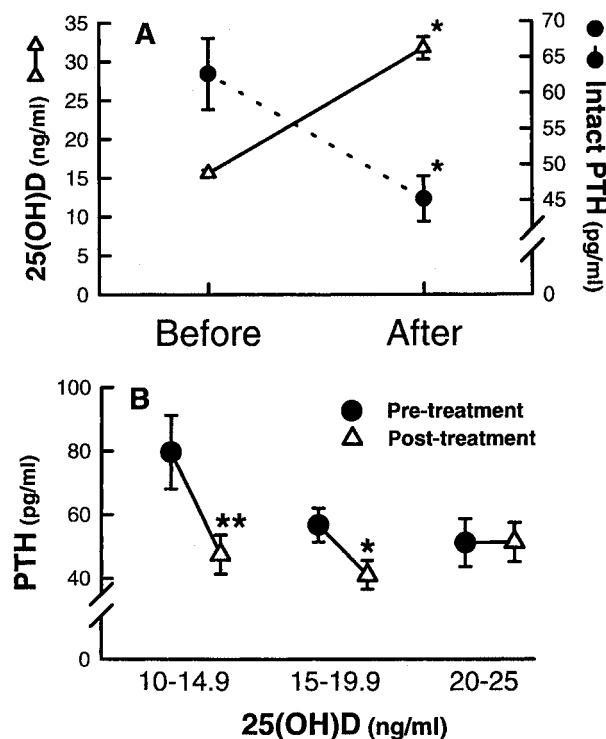


FIGURE 10. A: Serum concentrations of 25-hydroxyvitamin D [25(OH)D; ▲] and parathyroid hormone (PTH; ●) before and after therapy with 50 000 IU vitamin D₂ and calcium supplementation once a week for 8 wk. * $P < 0.05$. B: Serum concentrations of PTH in patients who had 25(OH)D concentrations between 10 and 25 ng/mL and who were stratified in increments of 5 ng/mL before (●) and after (▲) receiving 50 000 IU vitamin D₂ and calcium supplementation for 8 wk. * $P < 0.05$, ** $P < 0.01$. Reproduced with permission (71).

young adults with vitamin D deficiency were at greater risk of congestive heart failure than were their vitamin D-sufficient counterparts (73, 74).

Therefore, to maximize health and reduce the risk of common diseases, it is reasonable to pay attention to the 25(OH)D concentration. Just as the blood concentration of cholesterol is often measured on an annual basis, so too should the blood concentration of 25(OH)D be measured. Indeed, vigilance in maintaining a healthy 25(OH)D concentration may have more important health ramifications than a simple lowering of a blood cholesterol concentration to prevent coronary artery disease. A minimum concentration of 25(OH)D should be 50 nmol/L, and, for maximum bone health and prevention of many chronic diseases, the 25(OH)D concentration should be 78–100 nmol/L.

The simplest way to obtain vitamin D is from moderate exposure to sunlight. I recommend that exposure of hands, face and arms, or arms and legs to sunlight for a period equal to 25% of the time that it would take to cause a light pinkness to the skin (1 minimum erythral dose) is sufficient not only to satisfy the body's requirement, but also to make sufficient amounts of vitamin D to store in the body for use on rainy days and during times when sun exposure is inadequate to produce enough vitamin D in the skin. I have provided guidelines for the amount of sun exposure needed by people of all skin types to achieve their vitamin D requirement without significantly increasing the risk of skin damage and skin cancer (9, 39). Increasing the intakes of foods fortified with vitamin D, including milk, orange juice, cereals, and oily fish, is a reasonable approach to

satisfying the body's requirement. Taking > 1 multivitamin is counterproductive, because too much vitamin A would be ingested, and that increases the risk of birth defects and osteoporosis. Alternatively, one multivitamin containing 400 IU vitamin D and a vitamin D supplement containing either 400 or 1000 IU vitamin D is appropriate.

REFERENCES

- Holick MF. Phylogenetic and evolutionary aspects of vitamin D from phytoplankton to humans. In: Pang PKT, Schreibman MP, editors. Vertebrate endocrinology: fundamentals and biomedical implications. Vol 3. Orlando: Academic Press, 1989:7–43.
- Holick MF. Vitamin D. A millenium perspective. J Cell Biochem 2003; 88:296–307.
- Morris JG, Earle KE. Vitamin D and calcium requirements of kittens. Vet Clin Nutr 1996;3:93.
- Pitcher T, Pettifor JM, Buffenstein R. The effect of dietary calcium content and oral vitamin D₃ supplementation on mineral homeostasis in a subterranean mole-rat *Cryptomys damarensis*. Bone Miner 1994;27: 145–57.
- How KL, Hazewinkel HAW, Mol JA. Dietary vitamin D dependence of cat and dog due to inadequate cutaneous synthesis of vitamin D. Gen Comp Endocrinol 1994;96:12–8.
- Velluz L, Amiard G, Petit A. Le precalciferol—ses relations d'équilibre avec le calciferol. [Precalciferol and its relationship of equilibrium with calciferol.] Bull Soc Chim Fr 1949;16:501–8(in French).
- Holick MF, Tian XQ, Allen M. Evolutionary importance for the membrane enhancement of the production of vitamin D₃ in the skin of poikilothermic animals. Proc Natl Acad Sci U S A 1995;92:3124–6.
- Haddad JG, Matsuoka LY, Hollis BW, Hu YZ, Wortsman J. Human plasma transport of vitamin D after its endogenous synthesis. J Clin Invest 1993;91:2552–5.
- Holick MF. Vitamin D. The underappreciated D-lightful hormone that is important for skeletal and cellular health. Curr Opin Endocrinol Diabetes 2002;9:87–98.
- Matsuoka LY, Ide L, Wortsman J, MacLaughlin J, Holick MF. Sunscreens suppress cutaneous vitamin D₃ synthesis. J Clin Endocrinol Metab 1987;64:1165–8.
- Holick MF. McCollum Award Lecture, 1994: vitamin D—new horizons for the 21st century. Am J Clin Nutr 1994;60:619–30.
- Clemens TL, Henderson SL, Adams JS, Holick MF. Increased skin pigment reduces the capacity of skin to synthesis vitamin D₃. Lancet 1982;1:74–6.
- Webb AR, Kline L, Holick MF. Influence of season and latitude on the cutaneous synthesis of vitamin D₃; exposure to winter sunlight in Boston and Edmonton will not promote vitamin D₃ synthesis in human skin. J Clin Endocrinol Metab 1988;67:373–8.
- Lu Z, Chen TC, Kline L, et al. Photosynthesis of previtamin D₃ in cities around the world. In: Holick MF, Kligman A, eds. Biologic effects of light. Symposium proceedings, October 13–15, 1991. Berlin: Walter De Gruyter & Company, 1992:48–51.
- Tangpricha V, Koutkia P, Rieke SM, Chen TC, Perez AA, Holick MF. Fortification of orange juice with vitamin D: a novel approach to enhance vitamin D nutritional health. Am J Clin Nutr 2003;77:1478–83.
- MacLaughlin J, Holick MF. Aging decreases the capacity of human skin to produce vitamin D₃. J Clin Invest 1985;76:1536–8.
- Holick MF, Matsuoka LY, Wortsman J. Age, vitamin D, and solar ultraviolet. Lancet 1989;1:1104–5.
- Reid IR, Gallagher DJA, Bosworth J. Prophylaxis against vitamin D deficiency in the elderly by regular sunlight exposure. Age Aging 1985; 15:35–40.
- Chuck A, Todd J, Diffey B. Subliminal ultraviolet-B irradiation for the prevention of vitamin D deficiency in the elderly: a feasibility study. Photochem Photobiomed 2001;17:168–71.
- Specker B, Tsang RC, Hollis BW. Effect of race and diet on human-milk vitamin D and 25-hydroxyvitamin D. Am J Dis Child 1985;139:1134–7.
- Kreiter SR, Schwartz RP, Kirkman HN, Charlton PA, Calikoglu AS, Davenport M. Nutritional rickets in African American breast-fed infants. J Pediatr 2000;137:2–6.
- Nesby-O'Dell S, Scanlon KS, Cogswell ME, Gillespie C, Hollis BW, Looker AC. Hypovitaminosis D prevalence and determinants among African American and white women of reproductive age: third National

- Health and Nutrition Examination Survey, 1988–1994. *Am J Clin Nutr* 2002;76:187–92.
23. Tangpricha V, Pearce EN, Chen TC, Holick MF. Vitamin D insufficiency among free-living healthy young adults. *Am J Med* 2002;112:659–62.
 24. Bell NH, Epstein S, Greene A, Shary J, Oexmann MJ, Shaw S. Evidence for alteration of the vitamin D-endocrine system in obese subjects. *J Clin Invest* 1985;76:370–3.
 25. Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF. Decreased bioavailability of vitamin D in obesity. *Am J Clin Nutr* 2000;72:690–3.
 26. Holick MF. Sunlight and vitamin D: both good for cardiovascular health. *J Gen Intern Med* 2002;17:733–5.
 27. Holick MF. Sunlight “D”ilemma: risk of skin cancer or bone disease and muscle weakness. *Lancet* 2001;357:4–6.
 28. Glerup H, Mikkelsen K, Poulsen L, et al. Commonly recommended daily intake of vitamin D is not sufficient if sunlight exposure is limited. *J Intern Med* 2000;247:260–8.
 29. Glerup H, Mikkelsen K, Poulsen L, et al. Hypovitaminosis D myopathy without biochemical signs of osteomalacia bone involvement. *Calcif Tissue Int* 2000;66:419–24.
 30. Brown EM, Pollak M, Seidman CE, et al. Calcium-ion-sensing cell-surface receptors. *N Engl J Med* 1995;333:234–40.
 31. Holick MF. Vitamin D: photobiology, metabolism, mechanism of action, and clinical applications. In: Favus MJ, ed. *Primer on the metabolic bone diseases and disorders of mineral metabolism*. 5th ed. Washington, DC: American Society for Bone and Mineral Research, 2003:129–37.
 32. Juppner H, Kronenberg HM. Parathyroid hormone. In: Favus MJ, ed. *Primer on the metabolic bone diseases and disorders of mineral metabolism*. 5th ed. Washington, DC: American Society for Bone and Mineral Research, 2003:117–24.
 33. Garland CF, Garland FC, Shaw EK, Comstock GW, Helsing KJ, Gorham ED. Serum 25-hydroxyvitamin D and colon cancer: eight-year prospective study. *Lancet* 1989;1:1176–8.
 34. Garland FC, Garland CF, Gorham ED, Young JF. Geographic variation in breast cancer mortality in the United States: a hypothesis involving exposure to solar radiation. *Prev Med* 1990;19:614–2.
 35. Ahonen MH, Tenkanen L, Teppo L, Hakama M, Tuohimaa P. Prostate cancer risk and pre-diagnostic serum 25-hydroxyvitamin D levels (Finland). *Cancer Causes Control* 2000;11:847–52.
 36. Hanchette CL, Schwartz GG. Geographic patterns of prostate cancer mortality. Evidence for a protective effect of ultraviolet radiation. *Cancer* 1992;70:2861–9.
 37. Apperly FL. The relation of solar radiation to cancer mortality in North America. *Cancer Res* 1941;1:191–5.
 38. Grant WB. An estimate of premature cancer mortality in the U. S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* 2002;94:1867–75.
 39. Holick MF, Jenkins M. The UV advantage. New York: iBooks, Inc (in press).
 40. Hernán MA, Olek MJ, Ascherio A. Geographic variation of MS incidence in two prospective studies of US women. *Neurology* 1999;51:1711–8.
 41. Rostand SG. Ultraviolet light may contribute to geographic and racial blood pressure differences. *Hypertension* 1979;30:150–6.
 42. Grant WB. An ecologic study of the role of solar UV-B radiation in reducing the risk of cancer using cancer mortality data, dietary supply data and latitude for European countries. In: Holick MF, editor. *Biologic effects of light 2001* (proceedings of a symposium, Boston, MA). Boston: Kluwer Academic Publishing, 2002:267–6.
 43. Grant WB. An ecologic study of dietary and solar ultraviolet-B links to breast carcinoma mortality rates. *Cancer* 2002;94:272–81.
 44. Bodiwala D, Luscombe CJ, Liu S, Saxby M, French M, Jones PW. Prostate cancer risk and exposure to ultraviolet radiation: further support for the protective effect of sunlight. *Cancer Lett* 2003;192:145–9.
 45. DeLuca HF, Cantorna MT. Vitamin D: its role and uses in immunology. *FASEB J* 2001;15:2579–85.
 46. Stumpf WE, Sar M, Reid FA, et al. Target cells for 1, 25-dihydroxyvitamin D₃ in intestinal tract, stomach, kidney, skin, pituitary, and parathyroid. *Science* 1979;206:1188–90.
 47. Manolagas SC, Provvedini DM, Tsoukas CD. Interactions of 1, 25-dihydroxyvitamin D₃ and the immune system. *Mol Cell Endocrinol* 1985;43:113–22.
 48. Mathieu C, Adorini L. The coming of age of 1, 25-dihydroxyvitamin D₃ analogs as immunomodulatory agents. *Trends Mol Med* 2002;8:174–9.
 49. Feldman D, Zhao XY, Krishnan AV. Vitamin D and prostate cancer. *Endocrinology* 2000;141:5–9.
 50. Holick MF. Clinical efficacy of 1, 25-dihydroxyvitamin D₃ and its analogues in the treatment of psoriasis. *Retinoids* 1998;14:12–7.
 51. Kragballe K, Beck HI, Sogaard H. Improvement of psoriasis by a topical vitamin D₃ analogue (MC903) in a double-blind study. *Br J Dermatol* 1988;119:223–30.
 52. Gregori S, Giarratana N, Smirlando S, Uskokovic M, Adorini L. A 1 α , 25-dihydroxyvitamin D₃ analog enhances regulatory t-cells and arrests autoimmune diabetes in NOD mice. *Diabetes* 2002;51:1367–74.
 53. Cantorna MT, Hayes CE, DeLuca HF. 1, 25-Dihydroxycholecalciferol inhibits the progression of arthritis in murine models of human arthritis. *J Nutr* 1998;128:68–72.
 54. Cantorna MT, Hayes CE, DeLuca HF. 1, 25-Dihydroxyvitamin D₃ reversibly blocks the progression of relapsing encephalomyelitis, a model of multiple sclerosis. *Proc Natl Acad Sci U S A* 1996;93:7861–4.
 55. Mathieu C, Waer M, Laureys J, Rutgeerts O, Bouillon R. Prevention of autoimmune diabetes in NOD mice by 1, 25 dihydroxyvitamin D₃. *Diabetologia* 1994;37:552–8.
 56. Hyponen E, Laara E, Jarvelin M-R, Virtanen SM. Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *Lancet* 2001;358:1500–3.
 57. Krause R, Buhning M, Hopfenmuller W, Holick MF, Sharma AM. Ultraviolet B and blood pressure. *Lancet* 1998;352:709–10.
 58. Li Y, Kong J, Wei M, Chen ZF, Liu S, Cao LP. 1, 25-dihydroxyvitamin D₃ is a negative endocrine regulator of the renin-angiotensin system. *J Clin Invest* 2002;110:229–38.
 59. Adams JA, Clemens TL, Parrish JA, Holick MF. Vitamin D synthesis and metabolism after ultraviolet radiation of normal and vitamin D deficient subjects. *N Engl J Med* 1981;306:722–5.
 60. Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ. Human serum 25-hydroxycholecalciferol responses to extended oral dosing with cholecalciferol. *Am J Clin Nutr* 2003;77:204–10.
 61. Vieth R. Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *Am J Clin Nutr* 1999;69:842–56.
 62. Weisman Y, Harell A, Edelstein S, et al. 1, 25-dihydroxyvitamin D₃ and 24, 25-dihydroxyvitamin D₃ in vitro synthesis by human decidua and placenta. *Nature* 1979;281:317–9.
 63. Bikle DD, Nemanic MK, Gee E, Elias P. 1, 25-Dihydroxyvitamin D₃ production by human keratinocytes: kinetics and regulation. *J Clin Invest* 1986;78:557–66.
 64. Schwartz GG, Whitlatch LW, Chen TC, Lokeshwar BL, Holick MF. Human prostate cells synthesize 1, 25-dihydroxyvitamin D₃ from 25-hydroxyvitamin D₃. *Cancer Epidemiol Biomarkers Prev* 1998;7:391–5.
 65. Tangpricha V, Flanagan JN, Whitlatch LW, et al. 25-hydroxyvitamin D-1 α -hydroxylase in normal and malignant colon tissue. *Lancet* 2001;357:1673–4.
 66. Cross HS, Bareis P, Hofer H, Bischof MG, Bajna E, Kriwanek S. 25-Hydroxyvitamin D₃-1 α -hydroxylase and vitamin D receptor gene expression in human colonic mucosa is elevated during early cancerogenesis. *Steroids* 2001;66:287–92.
 67. Mawer EB, Hayes ME, Heys SE, et al. Constitutive synthesis of 1, 25-dihydroxyvitamin D₃ by a human small cell lung cell line. *J Clin Endocrinol Metab* 1994;79:554–60.
 68. Harnden D, Kumar R, Holick MF, DeLuca HF. Side chain metabolism of 25-hydroxy-[26, 27-¹⁴C] vitamin D₃ and 1, 25-dihydroxy-[26, 27-¹⁴C] vitamin D₃ in vivo. *Science* 1976;193:493–4.
 69. Holick MF. Vitamin D. In: *Dietary reference intakes for calcium, phosphorus, magnesium, vitamin D, and fluoride*. Washington, DC: Institute of Medicine. National Academy Press, 1997:250–87.
 70. Heaney RP. Long-latency deficiency disease: insights from calcium and vitamin D. *Am J Clin Nutr* 2003;78:912–9.
 71. Heaney RP, Dowell MS, Hale CA, Bendich A. Calcium absorption varies within the reference range for serum 25-hydroxyvitamin D. *J Am Coll Nutr* 2003;22:142–6.
 72. Koutkia P, Lu Z, Chen TC, Holick MF. Treatment of vitamin D deficiency due to Crohn's disease with tanning bed ultraviolet B radiation. *Gastroenterology* 2001;121:1485–8.
 73. Zittermann A, Schleithoff SS, Tenderich G, Berthold HK, Körfer R, Stehle P. Low vitamin D status: a contributing factor in the pathogenesis of congestive heart failure? *J Am Coll Cardiol* 2003;41:105–12.
 74. Malabanan A, Veronikis IE, Holick MF. Redefining vitamin D insufficiency. *Lancet* 1998;351:805–6.