42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61



Immunology Letters 00 (2002) 1-10



www.elsevier.com/locate/

Immunoregulation of CNS autoimmunity by helminth and mycobacterial infections

Diane L. Sewell, Emily K. Reinke, Laura H. Hogan, Matyas Sandor, Zsuzsa Fabry*

Department of Pathology, University of Wisconsin, 1300 University Avenue, Room 6330, Madison, WI 53706, USA

Abstract

10 11 12

13

14

15

16 17

18

19

20

2.1

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

The 'hygiene hypothesis' has been proposed to explain apparent increases in autoimmune disease and allergy in areas of the world with improved health care and sanitation. This hypothesis proposes that the lack of serious childhood infections impairs development of an appropriately educated immune response. Imbalance of Th1 and Th2 responses and lack of regulatory T-cell populations are two of many proposed potential mechanisms for immune failures such as autoimmunity and allergy. We summarize the literature evidence for the influence of infectious organisms on autoimmunity with focus on helminth and mycobacterial infections. We also demonstrate that Schistosoma mansoni ova pretreatment, Mycobacterium bovis (BCG) infection, and lyophilized Mycobacterium tuberculosis all modify the course of clinical disease in mice induced for experimental autoimmune encephalomyelitis (a mouse model for human multiple sclerosis (MS)). Our data supports the applicability of the hygiene hypothesis to CNS autoimmune disease. © 2002 Published by Elsevier Science B.V.

Keywords: Autoimmunity; Multiple sclerosis; Experimental autoimmune encephalomyelitis; Mycobacterium; Helminth; Schistosoma;

Immunoregulation

1. Introduction

In the natural environment, the human immune repertoire is constantly shaped by environmental exposures to infectious agents, resulting in the generation of memory T-cells capable of responding rapidly to antigenic re-stimulation and establishing a pre-existing immune status. Memory T-cell responses are also continually modulated by ongoing autoimmune triggers and infectious exposures. This paper will review the evidence for the influence of infection by pathogenic microorganisms on autoimmune disease. We will specifically highlight interactions between helminth and mycobacterial infections and the CNS autoimmune diseases, multiple sclerosis, and its mouse model, experimental autoimmune encephalomyelitis (EAE). It is clear that autoimmunity and infectious diseases do not occur in isolation. The outcome of noninfectious diseases is influenced both by the pre-existing immune status of the

2. EAE as a model of CNS autoimmunity

EAE is one of best-studied autoimmunity models, characterized by an autoimmune attack on CNS myelin mediated by neural autoantigen specific T helper cells [3]. It is currently the best available model for human multiple sclerosis [4]. In the induction of EAE, autoreactive T-cells are activated in the periphery of mice by subcutaneous injection of either crude spinal cord extracts or CNS antigens including myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG) or proteolipid protein (PLP) or their peptides. Activated autoreactive T-cells access the CNS, in the presence of competent antigen presenting cells, they are further activated and induce a local inflammatory response (Fig. 1). In most models, the T helper 1 (Th1) subset of T-cells has been implicated in the induction phase of EAE. Activation of myelin antigen reactive T-cells by antigen presenting cells (APC) that have been activated by exposure to mycobacterium in CFA favors matura-

individual and by exposures to infectious pathogens from the natural environment [1,2].

^{*} Corresponding author. Tel.: +1-608-265-8715; fax: +1-605-265-3301.

E-mail address: zfabry@facstaff.wisc.edu (Z. Fabry).

^{0165-2478/02/\$ -} see front matter © 2002 Published by Elsevier Science B.V.

PII: S0165-2478(02)00025-1

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

MECHANISM OF CNS AUTOIMMUNITY

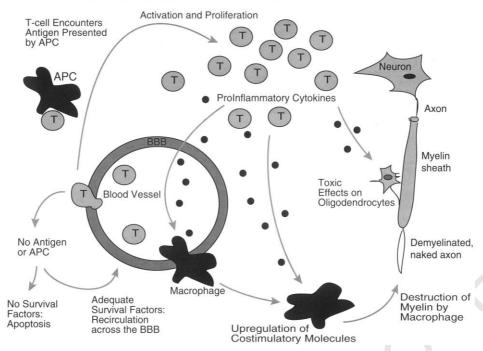


Fig. 1. Mechanism of CNS Autoimmunity. T-cells of the Th1 subset play a central role in CNS autoimmunity. These T-cells recirculate in and out of the CNS, providing surveillance. They can become activated by encounter with their cognate antigen or a molecular mimic, presented by APC's expressing costimulatory molecules, either inside the CNS or in the periphery. Activated T-cells produce pro-inflammatory cytokines including IFNγ. These cytokines act on the endothelium of the BBB enhancing transmigration of more T-cells and other inflammatory cells including macrophage. IFNγ also enhances expression of costimulatory molecules on APC's within the CNS, allowing more efficient presentation to T-cells, more activation and proliferation, potentiating the autoimmune pathology. Infiltrating macrophage phagocytose and present myelin peptides and produce $TNF\alpha$, a proinflammatory cytokine that has been shown to be toxic to oligodendrocytes in vitro [79]. Oligodendrocytes produce and maintain the myelin sheaths that insulate axons in the white matter of the CNS. The combination of myelin damage, phagocytosis and impaired repair result in clinical

tion of these Th1 cells. The mechanisms that lead to autoimmunity are still controversial, however in MS and its animal models, the role of autoimmune, functionally polarized Th 1 cells has been strongly suggested.

3. Th1 and Th2 subsets of CD4+ helper T-cells

At least two distinctly polarized subsets of antigenexperienced T-cells have been identified. Th1 cells secrete primarily IL-2, IFNγ, and TNF-β and express chemokine receptor CCR5 as well as IL18 receptor. Th2 cells produce IL- 4, -5, -6, -10 and IL-13 cytokines and express the G-protein linked receptors CCR3 and CCR4. Th1 cells influence the outcome of exposure to infectious pathogens and regulate autoimmune diseases, whereas Th2 cells are the key effectors in response to allergies and helminthic infections (Fig. 2) [5]. Th1 and Th2 clones also differ in their requirements for antigen presentation. Different antigen presenting cells, depending on their differentiation stage, activation status and the cytokine microenvironment, can preferentially stimulate T-cells to secrete Th1 or Th2 patterns of cytokines. The existence of functionally polarized human T-cell responses based on their profile of cytokine secretion has been established for both CD4⁺ T helper (Th) and CD8⁺ T cytotoxic cell subsets (Tc) [6].

The contributing role of different factors inducing Thelper cell differentiation into the polarized Th1 or Th2 pathway has been controversial, however, it has been demonstrated that infectious pathogens play an important role in this process. It is clear that there is a differential cytokine profile evoked by different infectious agents, influenced by the nature and concentration of the peptide ligand, the activity of co-stimulatory molecules, the local microenvironment of secreted hormones, and the context of different host genetic backgrounds. Moreover, polarized Th1-type and Th2type responses also play different roles in protection, with Th1 effective in the defense against intracellular pathogens and Th2 against intestinal nematodes. These different pathways are responsible for different types of immunopathologic reactions [6,17].

Infectious diseases have well-established effects on Th1/Th2 cytokine profiles. Mycobacterial infections are typically inducers of Th1 cytokines, IFNy, and lymphotoxin (TNFβ) [7]. Conversely, chronic parasitic infections such as schistosomiasis, and ascariasis induce

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

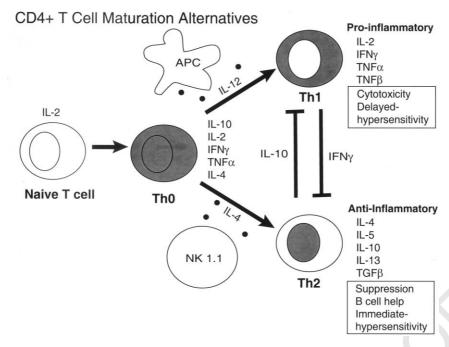


Fig. 2. CD4+ T-cell Maturation Alternatives. Naïve T-cells demonstrate potential to mature along two mutually exclusive pathways. The factors that determine the path a given T-cell will follow are not completely understood. The cytokine microenvironment plays a role with IL-12 encouraging maturation along the Th1 pathway and IL-4 encouraging Th2. Infectious agents dramatically influence T-helper maturation. Mycobacterium favors Th1 whereas extracellular parasites strongly favor Th2.

strongly Th2 polarized cytokine environments with predominant IL-4 and IL-5 [8–10]. The cytokine microenvironment also determines the maturation path of activated T-cells with IL-12 and IFN γ favoring Th1, and IL-4 and IL-10 favoring Th2 outcomes [11–16].

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

Although a highly cross-regulated control of polarized Th1 and Th2 cell types is a well-established paradigm, the exact mechanism of this cross talk is complex and remains under investigation. Cytokines produced by Th1 cells have negative regulatory effects on Th2 cells and vice versa. IFNy negatively impacts proliferation of Th2 cells [18] and IL-10 inhibits IFNy and other cytokine secretion by Th1 cells [19,20]. Most of the work describing cross-regulation of T helper subsets has been done in vitro using T-cell clones cultured in the presence of various cytokines and the result is observed. These models do not necessarily illuminate cross-regulatory events that take place in vivo [21]. There is plentiful evidence of cross-regulation in vivo however. Identification of a dichotomy in T-helper cells (Th1/Th2) helped explain observations of mutually exclusive DTH and antibody responses [22-24].

4. Infections, establishing a preexisting immune status in individuals, can modify the response to subsequent immune stimuli

The argument that development of the immune system is strongly influenced by continuous environ-

mental infectious stimulation and is capable of modifying subsequent immune responses has been supported by many observations. For example, patients infected with Schistosoma mansoni mount a Th2-type response to tetanus toxoid immunization instead of the more common Th1 or Th0 type response [9,25]. Furthermore, Ethiopian immigrants with a high prevalence of helminthic infections have eosinophilia and a propensity to respond to PHA with Th2-type, rather than Th1-type cytokines [26]. Infection of mice with S. mansoni delays clearance of vaccinia virus, an infection best controlled by a strong Th1 response. Mice develop a Th2 type response when infected with the microfilaria, Brugia malayi, or when immunized with soluble filarial extract from this parasite. The ongoing Th2 response to this helminth antigen modulates the Th1 response to nonparasite or microbial antigen [27,28]. Moreover, the murine intestinal nematode, Nippostrongylus brasiliensis stimulates Th2 activity. Rats infected with Nippostrongylus showed delays in kidney graft rejection (a DTH response), most likely by the cross-regulatory suppression of Th1 activity [29]. BCG vaccination cannot induce effective Th1 protection against tuberculosis in worm-infested areas. The efficacy of vaccination appears to be restored by treatment of helminthic infections [30].

We argue that these modified immune responses are not exclusively attributed to a modified Th1/Th2 response, but other factors are likely important in the establishment of a pre-existing immune status.

ARTICLE IN PRESS

D.L. Sewell et al. / Immunology Letters 00 (2002) 1-10

5. Infectious diseases frequently demonstrate a shift in T helper subset predominance in the natural course of the infection

4

163

164 165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199 200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

Measles infection initially induces a Th1 dominated response. Following clearance of the virus, the response shifts to a Th2 dominated response. The generalized immunosuppression following infection may result from viral infection of T and B-lymphocytes, PMNs and circulating monocytes and is responsible for the deaths from secondary infections that follow measles infection in developing countries. The role of B-cells as primary APC's may be responsible for the shift to a Th2 profile. It has also been theorized that Th2 cells are present from the beginning of the response and they are more longlived than Th1 cells [31]. Similar shifts from Th1 to Th2 responses have been observed in HIV infection [32] where the shift has been associated with onset of AIDS; in Plasmodium chabaudi chabaudi malaria (the mouse model for human falciparum malaria) where the life cycle of the parasite shifts from extracellular to intracellular; and in S. mansoni infection where the shift seems to correlate with the onset of egg laying by mature female worms [8].

Shifts from Th2 to Th1 have been observed less commonly, either occurring naturally or induced for therapeutic reasons. In a Leishmania model, Nabors et al. have been able to induce a therapeutic Th2 to Th1 shift in mice by administration of both Pentostam (a leismaniacidal drug) and IL-12. The mechanism is unclear, since normally, Th2 cells become unresponsive to IL-12 early in their differentiation [33].

6. Cross-reactive priming of autoimmune T-cells plays an important role in the induction of autoimmune disease

In spite of the generally accepted importance of autoimmune Th1 cells in the induction of autoimmunity, the exact mechanisms that lead to autoimmunity are still not clear. Infectious pathogens may play an important role in the initiation of autoimmune diseases. Activated, autoreactive T-cells can induce autoimmune disease whereas resting autoreactive T-cells cannot [34]. This has been demonstrated using several animal models of autoimmune disease; adoptive EAE, collagen induced arthritis (CIA) and herpes simplex keratitis (HSK). Pathogens have been implicated in the activation of normally innocuous, low affinity self-reactive T-cells [35,36]. The potential mechanisms for induction of autoimmunity by infectious agents are reviewed in more detail by Wucherpfennig[37]. Mechanisms by which infections may induce autoimmunity include molecular mimicry, superantigen activation of T-cells expressing targeted β chain alleles (V β), enhanced antigen processing by activated APC's, bystander activation, and activation of lymphocytes by lymphotropic viruses [37]. While infections are generally accepted as factors in induction of autoimmune disease, the focus of this review is on the lack of early exposure to helminth and mycobacterial infections as a risk factor in autoimmune disease.

7. Lack of early exposure to helminth and/or mycobacterial pathogens may be a risk factor for autoimmune disease

Autoimmunity and allergy are both on the rise worldwide, representing major concerns for the health-care system [38]. In the case of the increase in allergy, arguments have been made that the increase is related to a decrease in childhood infections as a result of improved sanitation and control of many previously endemic pathogens. The hygiene hypothesis suggests that there has been a population shift from T-helper 1 (Th1) to Th2 responses as a result of the cleaner environment [39–42]. One might have predicted a simple shift in the balance from Th1 to Th2 should have been accompanied by a concurrent decrease in autoimmune diseases that are predominately mediated by Th1 cells, i.e. MS, type 1 diabetes mellitus, inflammatory bowel disease (IBD) and others.

In fact, there has been a parallel increase in allergy and autoimmunity, both increasing predominantly in developed countries and in urban areas [38]. This pattern of concurrent increase of Th1 mediated and Th2 mediated diseases, both characterized perhaps by disordered immunoregulation, has led us and others to hypothesize that reduced exposure to both Th1-inducing and Th2-inducing pathogens in childhood can increase susceptibility to both allergy and autoimmunity [43]. In support of the hygiene hypothesis' applicability to autoimmune disease, there has been a recent report of an inverse relationship between risk of type 1 diabetes mellitus in children and daycare attendance and/or high numbers of contacts in early childhood [44]. Another report from Lithuania suggests that the occurrence of infection in the first 6 months of life correlates with lower incidence of type 1 diabetes and infection incidence at later times shows no correlation with diabetes incidence. The role of infections in the etiology of type 1 diabetes as well as other autoimmune diseases is controversial. Certain enteroviral infections might trigger the beta-cell destruction but insufficient exposure to early infections might increase the risk [45].

Here, we review data that support the link between a relative lack of infectious exposure and increased incidence of CNS and other autoimmune disease, and conversely, the protective effects of infectious agents in autoimmune diseases.

215 216

217 218 219

220

221

222

223

229

230

231

238

260261262

d 263 d 264 n 265 266

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

8. Mycobacteria prevent or ameliorate autoimmunity

267

268

269

270

271

272

273

274

275

276

277

278

279 280

281

282

283

284

285

286

287

288

289

290

291

292

293 294

295

296

297

298 299

300

301

302

303

304

305

306 307

308

309

310

311

312

313

314

315

316

317

318

319

Andersen et al. reported that a lack of exposure to both mycobacteria (as indicated by a negative tuberculin skin test) and measles (based on parental report at school enrollment) before age seven correlated with higher incidence of multiple sclerosis in adult life. This data was based on a retrospective case matched study including 92 MS patients and 276 age and sex matched controls selected from a reference population (births 1930-1950) of 198,000 school health records from Copenhagen, Denmark reported in 1981 [46]. We have addressed the role of mycobacterial infection in influencing autoimmunity in the CNS by infecting C57BL6 mice with Mycobacterium bovis strain BCG. Our data indicate that while MOG₍₃₅₋₅₅₎ peptide induces EAE in C57BL6 mice with 100% efficiency, BCG infected animals demonstrated a significant protection from this CNS autoimmune disease. When we infect these mice for 6 weeks with M. bovis strain BCG, they are protected from EAE as demonstrated by lower incidence, lower mean clinical scores and later onset (manuscript in preparation). We have also seen a therapeutic effect of intraperitoneally injected, lyophilized M. tuberculosis that is most dramatic when the bacteria are given at 2 days post induction of EAE. There was a significant improvement in clinical EAE when M. tuberculosis was given 4, 7 and 10 days post EAE induction. Thus, this treatment has some efficacy up to the time of onset of symptoms (Fig. 3).

The protective effect of mycobacterial components was reported in a guinea pig model of EAE as early as 25 years ago [47]. It has also been suggested that purified protein derivative (PPD) is the major component of M. tuberculosis implicated in protection from EAE. Recently, a 12-kDa PPD protein was demonstrated to be important in the protective activity of PPD [48,49]. Sequence studies indicated that this 12-kDa protein might belong to the bacterial heat shock protein family. Thus, similarly to hsp65-induced protection in arthritis or diabetes, the mechanism of protection might be based on shared T-cell epitopes with target self-antigen. Furthermore, Lehmann et al. reported that Bordetella pertussis is effective in inducing protection against EAE in SJL and SJLxBalbC F1 mice. According to their study, the mechanisms of the protective effects of Bordetella pertussis and mycobacterium in EAE are not the same. Adoptive transfer experiments indicated that the protection by M. tuberculosis is mediated by Tcells whereas similar transfers of B. pertussis sensitized T-cells are not protective [50]. Interestingly, both of these organisms have been routinely used for their adjuvant effects in the induction of EAE [48,49]. Brenner et al. demonstrated that protection from EAE

Modulation of EAE by Mycobacterium tuberculosis

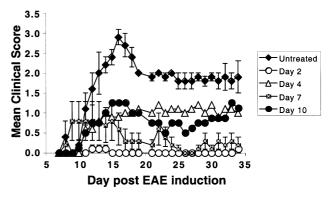


Fig. 3. Modulation of EAE by intraperitoneal injection of heat-killed M. tuberculosis. Six-week-old female C57BL/6 mice were induced for EAE by injection of 100 μg MOG₃₅₋₅₅ peptide emulsified in CFA that had been supplemented to 5 mg/ml with M. tuberculosis H37Ra. Pertussis toxin, 200 ng in 500 µl sterile PBS was given intraperitoneally on the day of EAE induction and again on day 2. Two hundred microliters of heat killed M. tuberculosis H37Ra (1 mg/ml) in sterile PBS was injected intraperitoneally at 2, 4, 7 or 10 days post induction of EAE or not. Mice that received mycobacterium treatment at 2 days post EAE induction showed the most dramatic protection from EAE. Mycobacterium treatment showed some efficacy even as late as day 10 post EAE induction. Data points are mean daily clinical score of five animals per group ± S.E.M. Error bars are shown on days 2, 7 and untreated groups only for clarity. Similar variance was seen in the other two groups.

by B. pertussis toxin immunization is mediated by antibodies. In a fostering experiment, they showed that protection is transferred to newborn rats during lactation. This protection could also be passively transferred by intraperitoneal injection of immune serum to naïve adult rats [51].

Mycobacterial exposure has shown protective effects in other, non-CNS autoimmune diseases. Non-obese diabetic mice develop insulin dependent diabetes mellitus with high incidence when not exposed to mycobacteria. M. avium infection induces resistance to diabetes in these mice. The authors report that the resistance induced by mycobacteria seems to be mediated by a Th1 subset consistent with a regulatory (CD45RB low, CD38⁺) population that triggers anergy or deletion of self-reactive peripheral lymphocytes [52,53]. Adjuvant arthritis (AA) also seems to be prevented or ameliorated by early exposure to mycobacteria. Lewis rats, intraperitoneally infected shortly after birth with BCG develop less severe AA than their uninfected littermates when AA is induced by a standard protocol [54]. We have summarized the evidence for bacterial infection induced protection from autoimmune disease from the literature and from our research in Table 1.

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

6

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

Organism	Autoimmune disease	Species	Reference
Mycobacterial cell wall components	EAE	Guinea pigs	[47]
B. pertussis	EAE	SJL mice	[50,51]
M. tuberculosis	EAE	SJL mice	[50]
M. bovis BCG	EAE	C57BL6 mice	Sewell et al., in preparation
M. avium	IDDM	NOD mice	[52,53]
M. bovis BCG	Adjuvant arthritis	Lewis rats	[54]

9. Evidence of early helminth exposure in modulation of CNS and other autoimmune diseases

In contrast to bacterial infections, helminth or parasitic infections are known to induce Th2-type immunity [8,55]. The correlation between these types of infections and lower incidence of autoimmune diseases has been suggested previously. For example, MS occurs very rarely in areas with endemic schistosome infections, as opposed to the higher incidence of MS and other autoimmune diseases in areas with more stringent hygienic standards [56,57]. This difference suggests an inverse correlation between higher hygienic standards and the development of Th1 type autoimmune diseases. To address the nature of Th1 and Th2 cross-regulation in a natural immune environment, we asked whether 'natural Th2 pre-conditioning' of experimental animals would influence the development of Th1 modulated autoimmunity in a CNS autoimmune disease such as

We induced a Th2 environment in SJL mice by intraperitoneal and subcutaneous S. mansoni ova immunization (manuscript submitted). We observed a significant protection from EAE in S. mansoni ova pre-immunized animals, indicating that parasitic infections can influence the course of a CNS autoimmune disease, and suggesting the importance of an experienced immune system in autoimmunity. As some intestinal helminthic infections induce minimal pathology, infection or treatment with helminth components might offer a new therapeutic option to prevent and/or ameliorate MS.

Individuals with predominant Th2 responses against egg antigens have less severe egg-associated morbidity than those with predominantly Th1 responses, thus a Th2 predisposition in dealing with helminth eggs is selectively advantageous to the human host. From an evolutionary perspective, people living in endemic areas with high prevalence of helminth infections might be positively selected because of their adaptively advantageous Th2 responses. More importantly, many helminth parasites can survive in the host for many years [58]. The long-term exposure to helminth antigens, beginning early in childhood, may have a deep impact on maturation of the host's immune system. Similar protection from development of insulin dependent diabetes mellitus by infection with S. mansoni or by injections of schistosome eggs has been reported in susceptible nonobese diabetic mice [59].

The inverse relationship between risk of type 1 diabetes mellitus in children and daycare attendance and/or high numbers of contacts in early childhood [44] could be due to differences in helminth infestation. Daycare centers and institutions support the transmission of many infections. The report correlating first half year of life infections with lowered incidence of IDDM also supports the application of the hygiene hypothesis to autoimmunity [45]. Neither of these reports is specific concerning what types of infections were encountered at higher incidence in the respective protected populations. We would speculate, however, that increased contact with other young children early in life would increase exposure of a full spectrum of infectious diseases.

Elliott et al. have suggested that lack of exposure to helminth infections in childhood may be a factor in the increasing incidence of Crohn's disease [60]. Crohn's disease is an autoimmune disease with detectable organspecific antibodies against the intestinal goblet cells and acinar cells of the exocrine pancreatic tissue [61]. The autoimmune inflammation causes cramping, diarrhea, and bloating. This autoimmune disease can also be manifested in autoimmune attacks on other target organs such as the eye. According to Mayer et al., an evolution of understanding of the etiology of inflammatory bowel diseases (IBD), ulcerative colitis and Crohn's disease has occurred in the past 30 years. In the 60s and 70s, IBD was considered to be an autoimmune disease in which there was a directed attack by humoral and cellular elements of the immune system against intestinal tissues. Since that time, there has been growing appreciation that defects in cellular immunity, not autoreactive in nature, may play a larger role in disease pathogenesis [62].

Another organ specific autoimmune disease, collageninduced arthritis (CIA), a mouse model for rheumatoid arthritis, is down regulated by infection with Trypanosoma brucei brucei in DA rats. This protective effect was most significant when the rats were infected with live trypanosomes before induction of CIA [63]. Daniel-Ribeiro and Zanini reported that natural and 'patho-

404

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

genic' autoantibodies are protective against malaria and conversely, infection with malaria may offer protection from autoimmune disease [64,65]. In the same vein, polyclonal immunoglobulins from malaria infected BALB/c mice have shown a therapeutic effect on a lupus-like syndrome in a lupus prone strain (NZBxNZW F1 mice) [66].

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

Table 2 summarizes evidence for protective effects of parasitic infection in both CNS autoimmunity and several other autoimmune diseases.

In summary, the results of these studies provide evidence in support of the idea that infection with helminths can modulate the development of Th1 diseases by influencing the cytokine environment of immune competent cells. A large block of evidence suggests that long-term infections with helminths in childhood might have deep impact on the maturation of Th1 and Th2 cells. Exposure to helminths and other parasites, such as malaria and trypanosomes, could be an important factor in influencing the development of Th1 cell-mediated autoimmune diseases in adulthood.

10. Mechanisms of immunoregulation by helminth and mycobacterial infections

In the next paragraphs, we summarize several of the mechanisms that have been proposed in the literature illustrating our current understanding with regard to the immunoregulation by helminth and mycobacterial infections. It is probable that multiple mechanisms contribute to different extents to produce the final result. It is also likely that other mechanisms, not understood at this time, might play a role.

The most commonly accepted mechanism for the immunoregulatory effects of infectious agents on CNS autoimmunity is cross talk between Th1 and Th2 subsets [67]. The cross talk described in the Falcone and Bloom study entails a preconditioned Th2 response to KLH antigen resulting in a Th2 microenvironment influencing the maturation of autoreactive T-cells and resultant protection from EAE. This mechanism has been called immune deviation and several investigators have achieved improved results in autoimmune disease models using immune deviation strategies [68,69] (Fig. 4A).

Recent data suggests the importance of other regulatory or suppressor mechanisms on both innate and adaptive elements of the peripheral and CNS immune systems (Fig. 4B). Th2 cytokines, particularly IL-10, have been demonstrated to inhibit macrophage activation with resulting suppression of IL-12 production and Th1 differentiation. TGFB acts directly on Th1 cells inhibiting their growth and proliferation [70]. Regulatory or suppressor T-cells can play an active role in suppressing other T-cells. When suppressor T-cells exist, transfer of these cells can transfer tolerance. Th1 suppressor T-cells have been described. These cells express CD4 and CD25 (IL-2Rα). Their suppression requires cell-cell contact and antigen specificity. It is independent of soluble factors as demonstrated by the inability of supernatants from these cells to mediate suppression [71]. Cell surface expression of TGFβ on CD4+ CD25+ regulatory T-cells has been described recently [72]. The finding of surface-bound TGFβ provides clarification for seemingly contradictory reports concerning the role or lack of role for soluble factors including TGFβ in suppression.

Possibly, infectious foci play a role in protection from autoimmune disease by sequestration or modification of trafficking of auto-reactive T-cells. We have observed that activated T-cells traffic to granulomas regardless of their antigenic specificity (manuscript in preparation). The autoreactive T-cells needed to trigger CNS disease may potentially be prevented from reaching threshold levels in the CNS by re-directed trafficking to other preexisting inflammatory sites by strong chemokine gradients and/or shared addressins (Fig. 4C).

Although control of autoimmunity via immune modulation offers a very desirable therapy, we also have to consider the side effects of such a therapy. Genain et al. demonstrated that immune deviation can increase concentrations of pathogenic autoantibodies and in some circumstances exacerbate autoimmune disease in a marmoset EAE model. Marmosets were tolerized by intraperitoneal administration of soluble rMOG and demonstrated early protection from EAE followed by late lethal complications. High levels of MOG specific autoantibodies were demonstrated in the tolerized marmosets. Autoantibody generation (Fig. 4D) was attributed to Th2 cytokine effects on B-cells, induction of the shift from low affinity IgM antibodies to high

Table 2 Parasites with protective effects in autoimmune disease

Organism	Autoimmune disease	Species	Reference
S. mansoni ova	EAE	SJL mice	Qing et al., submitted
T. brucei brucei	CIA	DA rats	[63]
Malaria	Lupus syndrome	NZBxNZW mice	[66]
T. trichuria	IBD	Human	[60]
S. mansoni live infection or ova	IDDM	NOD mice	[59]

7

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

522

523

524

525

526527

528

529

530

531

532

533

534

535

536

537

538

539

540

ARTICLE IN PRESS

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

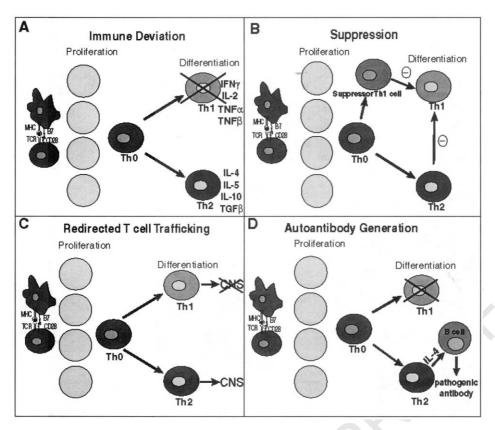


Fig. 4. Potential mechanisms for immune regulation of autoimmunity by helminth and mycobacterial infections. (A) Immune deviation is the suggested mechanism for helminth-induced protection from Th1 mediated autoimmune disease. Proliferation of T-cells in a microenvironment rich in IL-4 favors maturation along the Th2 pathway and suppresses the maturation of Th1 T-cells. (B) Chronic infections with mycobacteria and helminths have been suggested to induce suppressor cells by a variety of mechanisms. Suppressor cells downregulate inflammation and may control differentiation and expansion of the autoreactive T-cells that mediate autoimmunity. (C) Inflammatory foci that result from chronic mycobacterial and helminth infections may act as magnets for activated T-cells of many specificities. This could potentially keep the number of autoimmune T-cells that reach the target organ below the threshold required to induce disease. (D) Autoantibody generation is a potential late negative result of immune deviation. B-cells with the potential to make autoantibodies can be induced to proliferate, undergo affinity maturation and switch to IgG isotype in response to T-cell help and IL-4 cytokine.

affinity IgG1 [73]. Since both Th1 T-cells and myelin specific autoantibodies have been implicated in pathology of human MS, both T and B-cells need to be considered in any potential therapeutic regimen.

One common feature of microorganisms recognized by the immune system is the expression of pathogen associated molecular patterns (PAMPs) that are recognized by toll-like receptors (TLRs). These receptors are expressed on cells of the innate immune system [74,75]. The engagement of PAMPs with Toll-like receptors can also trigger the induction of IL-12 [76] and the induction of Th1 T-cell responses [77]. The role of PAMPs and TLRs in the regulation of innate immunity in the CNS can also influence adaptive immunity in the brain. These pathogens can also directly interact with APCs and modulate T-cell functions in this way. APC function can be altered not only by toll-like receptors, but also by other regulatory agents like cytokines. CD4 + T-cells can modify the capacity of APC's to induce autoimmune cells.

In summary, there are clearly multiple mechanisms providing immunoregulation by helminth and mycobacterial infections. To further understand these processes and how they interact will be crucial for our understanding of the complexity of immunoregulation.

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

11. Concluding remarks

Immunoregulation of CNS autoimmunity by mycobacterial pathogens has been reported previously [48–50]. Furthermore, it has also been suggested that mycobacterial infections play a beneficial role in allergic reactions [78]. In this paper, we argue the possibility of a more general paradigm of infectious pathogens as regulators of autoimmune reactions in the CNS. We summarize available data from the literature that suggests that not just mycobacteria, but also helminth pathogens are able to modulate CNS autoimmunity. Helminthic pathogens have also been demonstrated to

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

play a role in atopic diseases such as allergy [78]. 558 Confounding these observations, it is now indicated 559 that Th1-type autoimmunity can also be influenced by 560 561 parasites.

Further understanding of the regulatory mechanisms engaged by various classes of infectious pathogens on the immune system will aid our understanding of the extremely complex and enigmatic role they play in induction and prevention of CNS autoimmune diseases. The ultimate goal of this understanding would be to harness the immune regulatory effects of pathogenic organisms or their active components in control of CNS autoimmune diseases such as MS, without inducing the pathology that accompanies chronic infection with these organisms.

References

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

580

583 584

585

586

587

588

589

590

591

592

609

- [1] E.B. Bell, S.M. Sparshott, C. Bunce, Immunol. Today 19 (1998) 60 - 64.
 - [2] L.K. Selin, K. Vergilis, R.M. Welsh, S.R. Nahill, J. Exp. Med. 183 (1996) 2489-2499.
 - [3] H. Wekerle, K. Kojima, J. Lannes-Vieira, H. Lassmann, C. Linington, Ann. Neurol. 36 (1994) S47-S53.
 - [4] C.S. Raine, Lab. Invest. 50 (1984) 608-635.
- [5] S. Romagnani, M. Kapsenberg, A. Radbruch, L. Adorini, Res. 581 582 Immunol. 149 (1998) 871-873.
 - [6] S. Romagnani, Immunol. Today 12 (1991) 256-257.
 - [7] T. Mutis, Y.E. Cornelisse, T.H. Ottenhoff, Eur. J. Immunol. 23 (1993) 2189-2195.
 - E.J. Pearce, P. Caspar, J.M. Grzych, F.A. Lewis, A. Sher, J. Exp. Med. 173 (1991) 159-166.
 - [9] E.J. Pearce, A. La Flamme, E. Sabin, L.R. Brunet, Adv. Exp. Med. Biol. 452 (1998) 67-73.
 - [10] P.J. Cooper, M.E. Chico, C. Sandoval, I. Espinel, A. Guevara, M.W. Kennedy, J.F. Urban Jr, G.E. Griffin, T.B. Nutman, J. Infect. Dis. 182 (2000) 1207-1213.
- 593 [11] J.J. Bright, M. Rodriguez, S. Sriram, J. Virol. 73 (1999) 1637– 594
- 595 [12] C.S. Constantinescu, M. Wysocka, B. Hilliard, E.S. Ventura, E. 596 Lavi, G. Trinchieri, A. Rostami, J. Immunol. 161 (1998) 5097-597
- [13] E. Bettelli, M.P. Das, E.D. Howard, H.L. Weiner, R.A. Sobel, 598 599 V.K. Kuchroo, J. Immunol. 161 (1998) 3299-3306.
- 600 [14] M. Falcone, A.J. Rajan, B.R. Bloom, C.F. Brosnan, J. Immunol. 601 160 (1998) 4822-4830.
- 602 [15] J.J. Bright, B.F. Musuro, C. Du, S. Sriram, J. Neuroimmunol. 82 603 (1998) 22-30.
- [16] M.L. Krakowski, T. Owens, Eur. J. Immunol. 27 (1997) 2840-604 605
- [17] A.K. Abbas, M.E. Williams, H.J. Burstein, T.L. Chang, P. Bossu, 606 607 A.H. Lichtman, Immunol. Rev. 123 (1991) 5-22.
- 608 [18] T.F. Gajewski, E. Goldwasser, F.W. Fitch, J. Immunol. 141 (1988) 2635 - 2642.
- 610 [19] D.F. Fiorentino, M.W. Bond, T.R. Mosmann, J. Exp. Med. 170 611 (1989) 2081-2095.
- [20] T.R. Mosmann, K.W. Moore, Immunol. Today 12 (1991) A49-612 613 A 53
- 614 [21] P.A. Morel, T.B. Oriss, Crit. Rev. Immunol. 18 (1998) 275–303.
- 615 [22] C.R. Parish, F.Y. Liew, J. Exp. Med. 135 (1972) 298–311.
- 616 [23] C.R. Parish, Eur. J. Immunol. 2 (1972) 143-151.
- 617 [24] C.R. Parish, Transplant. Rev. 13 (1972) 35–66.

- [25] E.A. Sabin, M.I. Araujo, E.M. Carvalho, E.J. Pearce, J. Infect. Dis. 173 (1996) 269-272.
- [26] Z. Bentwich, Z. Weisman, C. Moroz, S. Bar-Yehuda, A. Kalinkovich, Clin. Exp. Immunol. 103 (1996) 239-243.
- [27] J.K. Actor, M. Shirai, M.C. Kullberg, R.M. Buller, A. Sher, J.A. Berzofsky, Proc. Natl. Acad. Sci. USA 90 (1993) 948-952.
- [28] M.C. Kullberg, E.J. Pearce, S.E. Hieny, A. Sher, J.A. Berzofsky, J. Immunol. 148 (1992) 3264-3270.
- [29] D.L. Ledingham, V.C. McAlister, H.N. Ehigiator, C. Giacomantonio, M. Theal, T.D. Lee, Transplantation 61 (1996) 184-188.
- [30] D. Elias, D. Wolday, H. Akuffo, B. Petros, U. Bronner, S. Britton, Clin. Exp. Immunol. 123 (2001) 219-225.
- [31] B.J. Ward, D.E. Griffin, Clin. Immunol. Immunopathol. 67 (1993) 171 - 177.
- [32] M. Clerici, G.M. Shearer, Immunol. Today 14 (1993) 107-111.
- [33] G.S. Nabors, L.C. Afonso, J.P. Farrell, P. Scott, Proc. Natl. Acad. Sci. USA 92 (1995) 3142-3146.
- [34] R.B. Fritz, X. Wang, M.L. Zhao, J. Neuroimmunol. 107 (2000) 66 - 72.
- [35] J. Thomas, B.T. Rouse, Immunol. Res. 16 (1997) 375-386.
- [36] S.P. Deshpande, S. Lee, M. Zheng, B. Song, D. Knipe, J.A. Kapp, B.T. Rouse, J. Virol. 75 (2001) 3077-3088.
- [37] K.W. Wucherpfennig, J. Clin. Invest. 108 (2001) 1097-1104.
- [38] P. Black, Trends Immunol. 22 (2001) 354-355.
- [39] G.A. Rook, J.L. Stanford, Immunol. Today 19 (1998) 113-116.
- [40] G. Folkerts, G. Walzl, P.J. Openshaw, Immunol. Today 21 (2000) 118 - 120
- [41] D.P. Strachan, Br. Med. J. 299 (1989) 1259-1260.
- [42] D.P. Strachan, Thorax 55 (Suppl. 1) (2000) S2-S10.
- [43] G.A. Rook, Immunol. Today 21 (2000) 249-250.
- [44] P.A. McKinney, M. Okasha, R.C. Parslow, G.R. Law, K.A. Gurney, R. Williams, H.J. Bodansky, Diabet. Med. 17 (2000) 236 - 242.
- [45] A. Pundziute-Lycka, B. Urbonaite, G. Dahlquist, Diabetologia 43 (2000) 1229 - 1234.
- [46] E. Andersen, H. Isager, K. Hyllested, Acta Neurol. Scand. 63 (1981) 131-135.
- [47] T.J. Meyer, I. Azuma, E.E. Ribi, Immunology 28 (1975) 219–229.
- [48] A. Ben-Nun, S. Yossefi, D. Lehmann, Eur. J. Immunol. 23 (1993) 689-696.
- [49] A. Ben-Nun, I. Mendel, G. Sappler, N. Kerlero de Rosbo, J. Immunol. 154 (1995) 2939-2948.
- [50] D. Lehmann, A. Ben-Nun, J. Autoimmun. 5 (1992) 675-690.
- [51] T. Brenner, O. Abramsky, J. Neurol. Sci. 114 (1993) 13-19.
- [52] T.C. Martins, A.P. Aguas, Clin. Exp. Immunol. 115 (1999) 248-
- [53] T.C. Martins, A.P. Aguas, Immunology 96 (1999) 600-605.
- [54] N. Esaguy, A.P. Aguas, Clin. Exp. Immunol. 104 (1996) 103–107.
- [55] E. Pearlman, J.W. Kazura, F.E. Hazlett, W.H. Boom, J. Immunol. 151 (1993) 4857-4864.
- [56] J.F. Kurtzke, Acta Neurol. Scand. Suppl. 161 (1995) 23-33.
- [57] J.F. Kurtzke, The Epidemiology of Multiple Sclerosis, Chapman and Hall, London, 1997.
- [58] A.E. Butterworth, D.W. Dunne, A.J. Fulford, K.J. Thorne, K. Gachuhi, J.H. Ouma, R.F. Sturrock, Immunol. Invest. 21 (1992)
- [59] A. Cooke, P. Tonks, F.M. Jones, H. O'Shea, P. Hutchings, A.J. Fulford, D.W. Dunne, Parasite Immunol. 21 (1999) 169-176.
- [60] D.E. Elliott, J.J. Urban, C.K. Argo, J.V. Weinstock, FASEB J. 14 (2000) 1848-1855.
- [61] J. Sykora, J. Varvarovska, F. Stozicky, M. Haschova, J. Hanzlikova, Cas. Lek. Cesk. 139 (2000) 735-737.
- [62] L. Mayer, Mt. Sinai J. Med. 67 (2000) 208-213.
- [63] L. Mattsson, P. Larsson, H. Erlandsson-Harris, L. Klareskog, R.A. Harris, Clin. Exp. Immunol. 122 (2000) 477-483.
- [64] C.T. Daniel-Ribeiro, Mem. Inst. Oswaldo. Cruz. 95 (2000) 199-207.

9

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

651 652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669 670

671

672

673

674

675

676

677

678

679

680

681

682

683

700

701

702

703

704

705

706

707

708

709

710

D.L. Sewell et al. | Immunology Letters 00 (2002) 1-10

685	[65] C.T. Daniel-Ribeiro, G. Zanini, Acta Trop. 76 (2000) 205-22
686	[66] B. Hentati, M.N. Sato, B. Payelle-Brogard, S. Avrameas,

10

687

688

689

690

691

692

693

694

695

696

697

698

- [66] B. Hentati, M.N. Sato, B. Payelle-Brogard, S. Avrameas, T. Ternynck, Eur. J. Immunol. 24 (1994) 8-15.
- [67] M. Falcone, B.R. Bloom, J. Exp. Med. 185 (1997) 901-907.
- [68] M.K. Racke, A. Bonomo, D.E. Scott, B. Cannella, A. Levine, C.S. Raine, E.M. Shevach, M. Rocken, J. Exp. Med. 180 (1994) 1961-1966.
- [69] M. Rocken, M. Racke, E.M. Shevach, Immunol. Today 17 (1996) 225 - 231.
- [70] Y. Chen, J. Inobe, V.K. Kuchroo, J.L. Baron, C.A. Janeway, Jr, H.L. Weiner, Proc. Natl. Acad. Sci. USA 93 (1996) 388-391.
- [71] A.M. Thornton, E.M. Shevach, J. Exp. Med. 188 (1998) 287-296.
- [72] K. Nakamura, A. Kitani, W. Strober, J. Exp. Med. 194 (2001) 629 - 644.

- [73] C.P. Genain, K. Abel, N. Belmar, F. Villinger, D.P. Rosenberg, C. Linington, C.S. Raine, S.L. Hauser, Science 274 (1996) 2054-
- [74] P. Brown, Nature 410 (2001) 1018-1020.
- [75] A. Aderem, R.J. Ulevitch, Nature 406 (2000) 782-787.
- [76] D.A. Hume, D.M. Underhill, M.J. Sweet, A.O. Ozinsky, F.Y. Liew, A. Aderem, BMC Immunol. 2 (2001) 11.
- [77] R.S. Chu, O.S. Targoni, A.M. Krieg, P.V. Lehmann, C.V. Harding, J. Exp. Med. 186 (1997) 1623-1631.
- [78] M. Yazdanbakhsh, A. van den Biggelaar, R.M. Maizels, Trends Immunol. 22 (2001) 372-377.
- [79] K.W. Selmaj, C.S. Raine, Ann. Neurol. 23 (1988) 339-346.